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(54) Title: GENE CONFERRING DISEASE RESISTANCE TO PLANTS (57) Abstract The present invention relates to an isolated gene fragment which confers disease resistance to plants by responding to an avirulence gene in plant pathogens. The gene fragment encodes for protein kinase, particularly serine/threonine kinase. The gene can be cloned into an expression vector to produce a recombinant DNA expression system suitable for insertion into cells to form a transgenic plant transformed with that gene fragment. Also disclosed is a process of conferring disease resistance to plants by growing plant host cells transformed with that expression system and expressing the gene conferring disease resistance to impart such resistance to the host cells.		

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GENE CONFERRING DISEASE RESISTANCE TO PLANTS

This invention arose out of research sponsored by NSF (Grant No. DMB-89-05997) and USDA/NRI (Grant No. 91-37300-6418).

FIELD OF THE INVENTION

The present invention relates to a gene conferring disease resistance to plants by responding to an avirulence gene in plant pathogens.

BACKGROUND OF THE INVENTION

Plants can be damaged by a wide variety of pathogenic organisms including viruses, bacteria, fungi and nematodes. Annual crop losses due to these pathogens is in the billions of dollars. Synthetic pesticides represent one form of defense against pathogens, and each year thousands of tons of such chemicals are applied to farm land and agricultural commodities. The cost of chemical pesticides is measured not only in the cost of producing these pesticides but also in both short term and long term environmental damage and the inherent risks to human health.

Plants also contain their own innate mechanisms of defense against pathogenic organisms. Natural variation for resistance to plant pathogens has been identified by plant breeders and pathologists and bred into many crop plants. These natural disease resistance genes often provide high levels of resistance (or immunity) to pathogens and represent the most economical and environmentally friendly form of crop protection. Despite the commercial significance, little is known about the molecular basis of natural disease resistance.

It has been postulated that disease resistance may be induced by the interaction of single genes in both the pathogen and the plant host. See A.H. Flor, "Host-Parasite Interactions in Flat-Rust -- its Genetics and Other Implications," Phytopath, 45:680-685 (1947) and A.H. Flor, "Current Status of the Gene-for-Gene Concept," Ann. Rev. Phytopath, 9:275-96 (1971), both of which are hereby

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incorporated by reference. Many plant disease resistance genes have been mapped to single loci, and individual avirulence genes have been isolated from bacterial and fungal pathogens. See B. Staskawicz, et al., "Cloned Avirulence Gene of *Pseudomonas syringae* pv. *glycinea* Determines Race-Specific Incompatibility on *Glycine max* Lo, Merr." Proc. Natl. Acad. Sci USA, 81:6024-28 (1984); A.M. Ellingboe, "Genetics of Host-Parasite Interactions," Encyclopedia of Plant Pathology, New Series, Vol. 4: Physiological Plant Pathology, pp. 761-78 (1976); D.W. Gabriel et al. "Gene-for-Gene Interactions of Five Cloned Avirulence Genes from *Xanthomonas campestris* pv *malvacearum* with Specific Resistance Genes in Cotton," Proc Natl Acad Sci USA 83:6415-19 (1986); S.H. Hulbert, et al., "Recombination at the *Rp1* Locus of Maize," Mol. Gen. Genet., 226:377-82 (1991); N.T. Keen, et al., "New Disease Resistance Genes in Soybean Against *Pseudomonas syringae* pv. *glycinea*: Evidence That One of Them Interacts with a Bacterial Elicitor," Theor. Appl. Genet. 81:133-38 (1991); R. Messeguer et al., "High Resolution RFLP Map Around the Root-knot Nematode Resistance Gene (*Mi*) in Tomato," Theor. Appl. Genet 82:529-3G (1991); T. Debener, et al., "Identification and Molecular Mapping of a Single *Arabidopsis thaliana* locus Determining Resistance to a Phytopathogenic *Pseudomonas syringae* isolate," Plant J 1:289:302 (1991); D.Y. Kobayashi, et al. "Cloned Avirulence Genes from Tomato Pathogen *Pseudomonas syringae* pv. *tomato* Confer Cultivar Specificity on Soybean," Proc. Natl. Acad. Sci. USA 86:157-61 (1989); and J.A.L. Van Kan, et al., "Cloning and Characterization of cDNA of Avirulence Gene *avr9* of the Fungal Pathogen *Cladosporium fulvum*, Causal Agent of Tomato Leaf Mold," Mol. Plant-Microbe Interactions 4:52-59 (1991), all of which are hereby incorporated by reference. However, despite this progress, the molecular isolation of plant disease resistance genes has been hindered by the fact that little is known of the gene products encoded at these loci.

The phenomenon of disease resistance is believed to be initiated by physical contact between a pathogen and a potentially compatible portion of the host. Once such contact

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has occurred, usually as a result of wind or rain vectored deposition of the pathogen, the pathogen must recognize that such contact has been established in order to initiate the pathogenic process. Likewise, such recognition by the host is required in order to initiate a resistance response. The precise manner in which such recognition occurs is not clear. However, pathogen recognition is believed to be associated with low pH of plant tissues or the presence of plant-specific metabolites. On the other hand, recognition by the host involves at least two partly separate pathways of recognition. A general mechanism detects a presence of the cell wall fragments from the pathogen and/or the damaged host. In addition, recognition results from a race-specific mechanism where the host disease resistance gene recognizes the avirulence gene of the pathogen. Both host recognition mechanisms lead to one or more levels of gene activation which in turn lead to production of defensive resistance factors (e.g., gum or cork production, production of inhibitors of pathogen proteases, deposition of lignin and hydroxyproplin-rich proteins in cell walls) and offensive resistance factors (e.g., production of phytoalexins, secreted chitinases). If the rate and level of activation of the genes producing these factors is sufficiently high, the host is able to gain an advantage on the pathogen. On the other hand, if the pathogen is fully activated at an earlier stage in the infection process, it may overwhelm both the offensive and defensive resistance factors of the plant. The phenomenon of disease resistance is fully discussed in J.L. Bennetzen et al., "Approaches and Progress in the Molecular Cloning of Plant Disease Resistance Genes," Genetic Engineering, 14:99-124 (1992), which is hereby incorporated by reference. Recently, elicitors of plant defense responses have been shown to induce phosphorylation and dephosphorylation of specific plant proteins, and inhibitors of mammalian protein kinases were found to inhibit expression of certain plant defense genes. See G. Felix, et al., "Rapid Changes of Protein Phosphorylation are Involved in Transduction of the Elicitor Signal in Plant Cells," Proc. Natl. Acad. Sci. USA, 88:8831-34

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(1991); V. Raz, et al., "Ethylene Signal is Transduced via Protein Phosphorylation Events in Plants," The Plant Cell, 5:523-30 (1993); and E.E. Farmer, et al. "Oligosaccharide Signaling in Plants -- Specificity of Oligouronide-Enhanced Plasma Membrane Protein Phosphorylation," J. Biological Chemistry, 266:3140-45 (1991), all of which are hereby incorporated by reference. At best, these references suggest that kinases are present in the metabolic pathway of disease resistance. These publications, however, do not disclose a gene which confers disease resistance to plants by responding to an avirulence gene in plant pathogens.

In tomato, resistance to the bacterial pathogen *Pseudomonas syringae* pv. *tomato* is encoded by a single locus (*Pto*) that displays dominant gene action. See R.E. Pitbaldo et al., "Genetic Basis of Resistance to *Pseudomonas syringae* pv. *tomato* in Field Tomatoes," Can. J. Plant Path., 5:251-55 (1983) ("Pitbaldo 1983"), which is hereby incorporated by reference. As with many commercially important traits in cultivated tomato (*Lycopersicon esculentum*), the resistance was identified in a wild tomato species, specifically *Lycopersicon pimpinellifolium*. See Pitbaldo 1983. Since the *Pto* gene was introgressed into tomato from a wild species, the region around the locus is polymorphic with respect to *L. esculentum* DNA. This polymorphism has been exploited by using a strategy relying on near-isogenic lines to identify molecular markers closely linked to *Pto*. See G.B. Martin, et al., "Rapid Identification of Markers Linked to *Pseudomonas* Resistance Gene in Tomato Using Random Primers and Near-isogenic Lines," Proc. Natl. Acad. Sci. USA, 88:2336-40 (1991) ("Martin et. al. 1991"), which is hereby incorporated by reference. Significant effort has been undertaken to map genetically the *Pto* gene. See G.B. Martin, et al. "High Resolution Linkage Analysis and Physical Characterization of the *Pto* Bacterial Resistance in Tomato," Molecular Plant Microbe Interaction, 6:21-34 (1993) ("Martin et. al. 1993") and G.B. Martin, et al, "Towards Positional Cloning of the *Pto* Bacterial Resistance Locus From Tomato," Advances in Molecular Genetics of Plant-Microbe Interactions, pp. 451-55 (1993).

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Moreover, the *Pto* gene is present in a number of commercial tomato varieties where it provides complete protection against *Pseudomonas syringae* pv. tomato bacteria and the disease referred to as "bacterial specks". Despite its wide-spread commercial use, no one has cloned or molecularly analyzed/characterized the *Pto* gene from tomato or a related disease resistance gene from any other plant species.

SUMMARY OF THE INVENTION

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The present invention relates to an isolated gene fragment which confers disease resistance to plants by responding to an avirulence gene in plant pathogens. It has been found that the gene fragment encodes for a protein kinase, more particularly a serine/threonine kinase. This gene can be inserted into an expression vector to produce a recombinant DNA expression system which forms another aspect to the present invention.

In another aspect of the present invention, a heterologous DNA conferring disease resistance to plants by responding to an avirulence gene in plant pathogens can be used to transform cells from transgenic plants. Again, the gene fragment encodes for protein kinase, particularly serine/threonine kinase. A process of conferring disease resistance to plants by growing plant host cells transformed with a recombinant DNA expression system comprising an expression vector into which this heterologous DNA is inserted and then expressing the heterologous DNA in the host cells to confer disease resistance is also disclosed.

In yet another aspect of the present invention, an isolated protein is disclosed which confers disease resistance to plants. That protein comprises an amino acid sequence for protein kinase, particularly serine/threonine kinase.

35

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a linkage map of a 20cM (i.e. centimorgan) region of tomato chromosome 5 developed from an F_2

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population segregating for *Pto*. This figure also shows the YAC clones identified with TG504, TG538, and TG475 as well as the 2 cDNAs identified by PTY 538-1. Additional markers at some loci are in parentheses.

5 Figure 2A shows an example of screening the YAC library with DNA markers.

Figure 2B shows a gel separation of a YAC clone using pulsed field gels. PTY538-1 is at 400kb. Another YAC (not relevant) was analyzed at 600kb.

10 Figure 3 shows genetic mapping of two end clones from PTY538-1.

Figure 4 shows genetic mapping of cDNA clones CD106 and CD127.

15 Figure 5 is a DNA blot analysis of YAC PTY538-1 (lane Y). Total yeast DNA from PTY538-1 was digested with *Bst*NI, separated on a 1% agarose gel and blotted onto Hybond N+ membrane. The membrane was probed with ³²P-labeled CD127 insert.

Figure 6 is a map of cloning vector lambda gt10.

20 Figure 7 is a map of plasmid vector pcDNAII. pcDNAII has a length of 3013 nucleotides, a first nucleotide of +1, a polylinker at bases 10-122, an *Sp6* promoter at bases 136-152, ampicillin resistance at bases 1331-2191, *F1* origin at bases 2377-2832, *Lac Z* gene at bases 2832-390, and a *T7* promoter at bases 2993-3012.

Figure 8 is a map of plasmid vector PTC3.

Figure 9 is a map of the T-DNA region of plasmid vector pPTC8.

30 Figure 10 is a map of the T-DNA region of plasmid vector pBI121.

Figure 11 is a plot of leaf bacteria versus time with the plotted value being means of 3 samples, each consisting of 3 leaf disks and error bars indicating standard deviations.

35 Figure 12 shows a DNA blot analysis of plant species distribution of *Pto* gene homologs.

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Figure 13A is a map of the CD186 cDNA insert, indicating the position and orientation of ORF1 (*Pto*) and 35S CaMV promoter on pPTC8.

Figure 13B is the deduced amino acid sequence for
 5 ORF1 (*Pto*) shown in standard 1-letter code.

Figure 14 is an RNA blot analysis of *Pto* gene family transcripts. PolyA+RNA of Rio Grande (*pto/pto*; lane 1), Rio Grande-*PtoR* (*Pto/Pto*; lane 2), Spectrum 151 (*pto/pto*; lane 3) and Moneymaker (*pto/pto*; lane 4) was isolated from leaf tissue
 10 of 5 week-old plants, separated on a 1.4% agarose-formaldehyde gel and blotted onto nitrocellulose. The blot was hybridized with ³²P-labeled CD127 insert. The difference in signal among samples is due to unequal loading of RNA as indicated by hybridizing the identical filter with a probe for ribulose
 15 biphosphate carboxylase transcript. Markers are RNA ladder (Gibco BRL, Gaithersburg, MD).

DETAILED DESCRIPTION OF THE INVENTION

20 The present invention relates to an isolated gene fragment conferring disease resistance to plants by responding to an avirulence gene in plant pathogens. The gene fragment encodes for protein kinase, particularly serine/threonine kinase. An amino acid sequence encoded by the gene fragment
 25 is SEQ. ID. No. 1 as follows:

Met	Gly	Ser	Lys	Tyr	Ser	Lys	Ala	Thr	Asn	Ser	Ile	Asn	Asp	Ala	Leu	1	5	10	15
Ser	Ser	Ser	Tyr	Leu	Val	Pro	Phe	Glu	Ser	Tyr	Arg	Val	Pro	Leu	Val	20	25	30	
Asp	Leu	Glu	Glu	Ala	Thr	Asn	Asn	Phe	Asp	His	Lys	Phe	Leu	Ile	Gly	35	40	45	
His	Gly	Val	Phe	Gly	Lys	Val	Tyr	Lys	Gly	Val	Leu	Arg	Asp	Gly	Ala	50	55	60	
Lys	Val	Ala	Leu	Lys	Arg	Arg	Thr	Pro	Glu	Ser	Ser	Gln	Gly	Ile	Glu	65	70	75	80
Glu	Phe	Glu	Thr	Glu	Ile	Glu	Thr	Leu	Ser	Phe	Cys	Arg	His	Pro	His	85	90	95	

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Leu Val Ser Leu Ile Gly Phe Cys Asp Glu Arg Asn Glu Met Ile Leu
 100 105 110
 Ile Tyr Lys Tyr Met Glu Asn Gly Asn Leu Lys Arg His Leu Tyr Gly
 115 120 125
 Ser Asp Leu Pro Thr Met Ser Met Ser Trp Glu Gln Arg Leu Glu Ile
 130 135 140
 Cys Ile Gly Ala Ala Arg Gly Leu His Tyr Leu His Thr Arg Ala Ile
 145 150 155 160
 Ile His Arg Asp Val Lys Ser Ile Asn Ile Leu Leu Asp Glu Asn Phe
 165 170 175
 Val Pro Lys Ile Thr Asp Phe Gly Ile Ser Lys Lys Gly Thr Glu Leu
 180 185 190
 Asp Gln Thr His Leu Ser Thr Val Val Lys Gly Thr Leu Gly Tyr Ile
 195 200 205
 Asp Pro Glu Tyr Phe Ile Lys Gly Arg Leu Thr Glu Lys Ser Asp Val
 210 215 220
 Tyr Ser Phe Gly Val Val Leu Phe Glu Val Leu Cys Ala Arg Ser Ala
 225 230 235 240
 Ile Val Gln Ser Leu Pro Arg Glu Met Val Asn Leu Ala Glu Trp Ala
 245 250 255
 Val Glu Ser His Asn Asn Gly Gln Leu Glu Gln Ile Val Asp Pro Asn
 260 265 270
 Leu Ala Asp Lys Ile Arg Pro Glu Ser Leu Arg Lys Phe Gly Asp Thr
 275 280 285
 Ala Val Lys Cys Leu Ala Leu Ser Ser Glu Asp Arg Pro Ser Met Gly
 290 295 300
 Asp Val Leu Trp Lys Leu Glu Tyr Ala Leu Arg Leu Gln Glu Ser Val
 305 310 315 320
 Ile

As demonstrated in the examples infra, a comparison of this
 sequence with those on available databases indicates that this
 sequence includes 11 subdomains, including 15 invariant amino
 acids, characteristic of protein kinases. In addition, there
 5 are sequences indicative of serine/threonine kinases.

Preferably, the gene fragment conferring disease
 resistance has nucleotide sequence SEQ. ID. No. 2 as follows:

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ATGGAAGCAAGTATTCTAAGGCAACAAATTCATATAATGATGCTTTAAGCTCGAGTTATCTCGT
 TCCTTTTGAAAGTTATCGAGTTCCTTTAGTAGATTTGGAGGAAGCAACTAATAATTTTGATCACA
 AGTTTTTAATTGGACATGGTGTCTTTGGGAAGGTTTACAAGGGTGTTTTGCGTGATGGAGCAAAG
 GTGGCCCTGAAAAGGCGTACACCTGAGTCCTCACAAAGGTATTGAAGAGTTCGAACAGAAATTGA
 5 GACTCTCTCATTTTGCAGACATCCGCATCTGGTTTCATTGATAGGATTCTGTGATGAAAGAAATG
 AGATGATTCTAATTTATAAATACATGGAGAATGGGAACCTCAAGAGACATTTGTATGGATCAGAT
 CTACCCACAATGAGCATGAGCTGGGAGCAGAGGCTGGAGATATGCATAGGGGCAGCCAGAGGTCT
 ACACTACCTTCATACTAGAGCAATTATACATCGTGATGTCAAGTCTATAAACATATTGCTTGATG
 AGAATTTTGTGCCAAAATTACTGATTTTGAATATCCAAGAAAGGGACTGAGCTTGATCAAACC
 10 CATCTTAGCACAGTAGTGAAAGGAACTCTCGGCTACATTGACCCTGAATATTTTATAAAGGGACG
 ACTCACTGAAAAATCTGATGTTTATTCTTTTCGGTGTGTTTTATTTCGAAGTCTTTGTGCTAGGT
 CTGCCATAGTTCAATCTCTTCCAAGGGAGATGGTTAATTTAGCTGAATGGGCAGTGGAGTCGCAT
 AATAATGGACAGTTGGAACAAATCGTAGATCCCAATCTTGCAGATAAAATAAGACCAGAGTCCCT
 CAGGAAGTTTGGAGATACAGCGGTAAATGCTTAGCTTTGTCTAGTGAAGATAGGCCATCAATGG
 15 GTGATGTGTTGTGGAACTGGAGTATGCACCTTCGTCTCCAAGAGTCTGTTATTTAA

The DNA molecule or gene fragment conferring disease
 resistance to plants by responding to an avirulence gene in
 plant pathogens can be incorporated in plant or bacterium
 20 cells using conventional recombinant DNA technology.
 Generally, this involves inserting the DNA molecule into an
 expression system to which the DNA molecule is heterologous
 (i.e., not normally present). The heterologous DNA molecule
 is inserted into the expression system or vector in proper
 25 orientation and correct reading frame. The vector contains
 the necessary elements for the transcription and translation
 of the inserted protein-coding sequences. A large number of
 vector systems known in the art can be used, such as,
 plasmids, bacteriophage virus or other modified viruses.
 30 Suitable vectors include, but are not limited to the following
 viral vectors such as lambda vector system λ gt11, λ gt10,
 Charon 4, and plasmid vectors such as pBI121, pBR322,
 pACYC177, pACYC184, pAR series, pKK223-3, pUC8, pUC9, pUC18,
 pUC19, pLG339, pRK290, pKC37, pKC101, pCDNAII, and other
 35 similar systems. The DNA sequences are cloned into the vector
 using standard cloning procedures in the art, as described by
 Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold
 Springs Laboratory, Cold Springs Harbor, New York (1982),
 which is hereby incorporated by reference. The plasmid pBI121

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is available from Clontech Laboratories, Palo Alto, California, (see Figure 9) has been used.

In order to obtain efficient expression of the gene or gene fragment of the present invention, a promoter must be present in the expression vector. RNA polymerase normally binds to the promoter and initiates transcription of a gene or a group of linked genes and regulatory elements (operon) in bacteria. Promoters vary in their strength, i.e., ability to promote transcription. Depending upon the host cell system utilized, any one of a number of suitable promoters can be used. Suitable promoters include nos promoter, the small subunit ribulose biphosphate carboxylase genes, the small subunit chlorophyll A/B binding polypeptide, the 35S promoter of cauliflower mosaic virus, and promoters isolated from plant genes, including the Pto promoter itself. See C.E. Vallejos, et al., "Localization in the Tomato Genome of DNA Restriction Fragments Containing Sequences Homologous to the rRNA (45S), the major chlorophyll a/b Binding Polypeptide and the Ribulose Biphosphate Carboxylase Genes," Genetics 112: 93-105 (1986) which discloses the small subunit materials. The nos promoter and the 35S promoter of cauliflower mosaic virus are well known in the art.

Once the disease resistance gene of the present invention has been cloned into an expression system, it is ready to be transformed into a plant cell. Plant tissue suitable for transformation include leaf tissues, root tissues, meristems, and protoplasts. It is particularly preferred to utilize explants of hypocotyls and cotyledons.

One technique of transforming plants with the gene conferring disease resistance in accordance with the present invention is by contacting the tissue of such plants with an inoculum of a bacteria transformed with a vector comprising a gene in accordance with the present invention which confers disease resistance and encodes for protein kinase. Generally, this procedure involved inoculating the plant tissue with a suspension of bacteria and incubating the tissue for 48 to 72 hours on regeneration medium without antibiotics at 25-28°C.

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Bacteria from the genus *Agrobacterium* can be utilized to transform plant cells. Suitable species of such bacterium include *Agrobacterium tumefaciens* and *Agrobacterium rhizogenes*. *Agrobacterium tumefaciens* (e.g., strains LBA4404 or EHA105) is particularly useful due to its well-known ability to transform plants.

Another approach to transforming plant cells with a gene which confers disease resistance and encodes for a protein kinase involves propelling inert or biologically active particles at plant tissues cells. This technique is disclosed in U.S. Patent Nos. 4,945,050, 5,036,006, and 5,100,792 all to Sanford et al., which are hereby incorporated by reference. Generally, this procedure involves propelling inert or biologically active particles at the cells under conditions effective to penetrate the outer surface of the cell and to be incorporated within the interior thereof. When inert particles are utilized, the vector can be introduced into the cell by coating the particles with the vector containing the gene conferring disease resistance and encoding for protein kinase. Alternatively, the target cell can be surrounded by the vector so that the vector is carried into the cell by the wake of the particle. Biologically active particles (e.g., dried yeast cells, dried bacterium or a bacteriophage, each containing DNA sought to be introduced) can also be propelled into plant cell tissue.

The isolated gene fragment of the present invention or related protein kinase genes can be utilized to confer disease resistance to a wide variety of plant cells, including gymnosperms, monocots, and dicots. Although the gene can be inserted into any plant cell falling within these broad classes, it is particularly useful in crop plant cells, such as rice, wheat, barley, rye, corn, potato, carrot, sweet potato, bean, pea, chicory, lettuce, cabbage, cauliflower, broccoli, turnip, radish, spinach, asparagus, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, quince, melon, plum, cherry, peach, nectarine, apricot, strawberry, grape, raspberry, blackberry, pineapple, avocado, papaya, mango, banana, soybean, tobacco,

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tomato, sorghum and sugarcane. The amino acid sequence identified by SEQ. ID. No. 1, and the nucleotide sequence, identified by SEQ. ID. No. 2, are particularly useful in conferring disease resistance to otherwise disease-prone
5 tomato plants. The present invention may also be used in conjunction with non-crop plants, including *Arabidopsis thaliana*.

The expression system of the present invention can be used to transform virtually any crop plant cell under
10 suitable conditions. Cells transformed in accordance with the present invention can be grown *in vitro* in a suitable medium to confer disease resistance by producing protein kinases. This protein can then be harvested or recovered by conventional purification techniques. The isolated protein
15 can be applied to plants (e.g., by spraying) as a topical application to impart disease resistance. Alternatively, transformed cells can be regenerated into whole plants such that this protein imparts disease resistance to the intact transgenic plants. In either case, the plant cells
20 transformed with the recombinant DNA expression system of the present invention are grown and caused to express that DNA in the cells to confer disease resistance on them.

Regardless of whether the DNA molecule of the present invention is expressed in intact plants or in culture,
25 expression of the desired protein follows essentially the same basic mechanism. Specifically, transcription of the DNA molecule is initiated by the binding of RNA polymerase to the DNA molecule's promoter. During transcription, movement of the RNA polymerase along the DNA molecule forms messenger RNA.
30 As a result, the DNA molecule that encodes for the hybrid protein of the present invention is transcribed into the corresponding messenger RNA. This messenger RNA then moves to the ribosomes of the rough endoplasmic reticulum which, with transfer RNA, translates the messenger RNA into the protein
35 conferring disease resistance to plants of the present invention. This protein then proceeds to trigger the plant's disease resistance mechanism. Although the sequence of events involved in the resistance mechanism is not well understood,

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it is expected that isolation of the gene fragment of the present invention and identification of its sequence will lead to a greater understanding of how disease resistance is conferred.

5

EXAMPLES

Example 1 - Identifying cDNA clones by map-based cloning.

- 10 A. High resolution linkage mapping and physical mapping of the Pto region on tomato chromosome 5.

Plant material and segregating populations

- 15 An F₂ population (86T64) derived from an interspecific cross of *Lycopersicon esculentum* cv. VF36-Tm2a x *L.pennellii* LA716 was used initially to assign markers to the Pto region. To order the markers with respect to Pto, F₂ population (90GM251) derived from a cross between two near-
20 isogenic lines differing for Pst susceptibility was used. Rio Grande-PtoR derives its resistance from *L.pimpinellifolium* and has undergone six backcrosses to Rio Grande and a final selfing generation.

- 25 *Scoring plant reactions to Fenthion and to Pseudomonas syringae pv. tomato*

- Approximately 100 seeds were sown per flat in the greenhouse (20-25OC) in a 1:2:1 (vol/vol) mixture of peat,
30 loam and perlite. Six weeks after germination, the plants were sprayed with a solution of 0.15% Fenthion/0.05% Silwet L-77 dispersed in sterile distilled water (Silwet L-77 source: Union Carbide, Southbury, CT.); Fenthion source: Mobay Corp., Kansas City, MO). After 3-4 days, small necrotic lesions (1-2
35 mm) were visible on controls known to be either homozygous or heterozygous at the Pto locus. A razor blade was used to cull all F₂ seedlings showing the necrotic lesions and one week later the Fenthion treatment and culling was repeated. Those seedlings which remained symptom-free after two Fenthion

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treatments were placed in the field and scored with flanking RFLP markers to identify plants having crossover events in the Pto region.

F₃ progeny from F₂ plants having crossovers near the Pto locus were screened for their reaction to *Pseudomonas syringae* pv. tomato (Pst; strain PT11) in the greenhouse as described by Martin et al. (1991), which is hereby incorporated by reference, except that instead of using cotton swabs the plants were dipped in a solution of 10⁶ colony-forming units per ml Pst strain PT11/0.05% Silwet L-77/10 mM MgCl₂ dispersed in sterile distilled water. Between 20 and 30 F₃ plants were sown as described above and treated at the three-to four-leaf stage. Reaction to the pathogen was scored after 7 days as either susceptible -- indicated by numerous necrotic specks surrounded by chlorotic halos -- or as resistant -- indicated by the absence of necrotic specks on the inoculated leaves.

Pulsed field gel electrophoresis

20

Preparation of tomato protoplasts (cvs. Rio Grande-PtoR and Rio Grande), isolation of high molecular weight DNA and digestion in agarose blocks was performed as described in M.W. Ganai et. al., "Analysis of Tomato DNA by Pulse Field Gel Electrophoresis," Plant Mol. Biol. Rep. 7:17-27 (1989) ("Ganai et. al. 1989"), which is hereby incorporated by reference. CHEF gels (Chu et al, 1986, which is hereby incorporated by reference) were used to separate the digested high molecular weight DNA. Gels were prepared in 0.5X TBE (1X TBE = 0.089 M Tris, 0.089 M boric acid, 0.002 M EDTA) at an agarose concentration of 1%. For DNA blotting, the gels were treated with UV light (254 nm for 5 minutes using a Fotodyne Transilluminator Model 3-4400) and then blotted onto Hybond N+ (Amersham Co.) using the recommended alkaline (0.4 N NaOH) blotting procedure. The molecular weight size standards included lambda concatamers (48.5 kb ladder, FMC Bioproducts, Rockland, ME) and yeast chromosomes from strain AB1380 (Burke et al. 1987), which is hereby incorporated by reference.

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Genetic linkage analysis

A linkage map for a cross between *L.esculentum* and
L.pennelli was constructed using Mapmaker software on a Sun II
5 workstation as described previously (Lander et al., 1987;
Tanksley et al., 1992, which are hereby incorporated by
reference). All markers shown placed with a LOD score of >3.
The ordering of the markers for the other two populations were
determined using Mapmaker and the ripple command gave a Δ LOD
10 of >2.9 for all alternative triple point placements.
Recombination frequencies between markers were calculated
manually using the maximum likelihood estimators of Allard
(1956), which is hereby incorporated by reference.

15 *Identification of markers in the Pto region*

Three approaches were used to identify a total of 28
markers, shown below in Table 1, linked to the *Pto* gene.

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TABLE 1

Marker	Type	Enzyme(s)**** showing RFLP between NILs	Reference, all of which are hereby incorporated by reference
CD31A	cDNA	EV, X	Bernatzky and Tanksley, 1986*
CT63A	cDNA	none	Tanksley et al., 1992**
CT104A	cDNA	none	Martin et al., 1993
CT155	cDNA	H	Tanksley et al., 1992
CT201A	cDNA	none	Tanksley et al., 1992
CT202	cDNA	none	Tanksley et al., 1992
CT260A	cDNA	B, X	Tanksley et al., 1992
R11	RAPD	E, EV	Martin et al., 1993
R53	RAPD	B, D, E, EV, H, X	Martin et al., 1993
R110	RAPD	B, E	Martin et al., 1991
RS120	RAPD	E, H	Martin et al., 1991
TG96	Sheared genomic	E, EV, X	Tanksley et al., 1992
TG100	Sheared genomic	B, H	Tanksley et al., 1992
TG318	Pst genomic	none	Tanksley et al., 1992
TG344B	Pst genomic	H	Martin et al., 1993
TG358	Pst genomic	none	Tanksley et al., 1992
TG379	Pst genomic	none	Tanksley et al., 1992
TG448	Pst genomic	none	Tanksley et al., 1992
TG475	Pst genomic	D, H,	Martin et al., 1993
TG478B	Pst genomic	none	Tanksley et al., 1992
TG503	Pst genomic	none	Tanksley et al., 1992
TG504	Pst genomic	B	Martin et al., 1993
TG538	Pst genomic	B, D, E, EV, H, X	Martin et al., 1993
TG606	Pst genomic	B, D, E, RV	Martin et al., 1993
TG619	Pst genomic	E, X	Tanksley et al., 1992
TG638	Pst genomic	EV	Martin et al., 1993
TM5	MADS Box gene	B, D, E, H, X	Pnueli et al., 1991***

*R. Bernatzky et. al., "Towards a Saturated Linkage Map in Tomato Based on Isozymes a cDNA clones," Genetics, 112:887-98 (1986), which is hereby incorporated by reference.

**S.D. Tanksley et. al., "High Density Molecular Linkage Maps of the Tomato and Potato Genomes," Genetics, 132:1141-60 (1992), which is hereby incorporated by reference.

***Pnueli et. al., "The MADS Box Gene Family in Tomato: Temporal Expression During Floral Development, Conserved Secondary Structures and Homology with Homeotic Genes from *Antirrhinum* and *Arabidopsis*," Plant J., 1:255-66 (1991), which is hereby incorporated by reference.

**** X=XbaI
E=EcoRI
D=DraI
Ev=EcoRV
B=BstNI
H=HaeIII

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First, a genome-wide mapping effort placed 1000 RFLP markers on the tomato map and identified 19 markers in the general *Pto* region which were then surveyed on the *Pto* near-isogenic lines (NILs) to identify informative clones. Secondly, surveys of
5 the resistant and susceptible NILs were probed with pools of 5 random clones (600 total clones) to identify polymorphic probes. Finally, RAPD analysis using 150 primers of arbitrary sequence (each amplifying about 4 products) was used to identify additional linked markers. The markers identified by
10 the latter two approaches were initially placed on the whole genome map (population 86T64) to confirm their placement to the *Pto* region. Each marker was then hybridized to survey filters of NILs DNA digested with six restriction enzymes to detect the general level of polymorphism exhibited by the
15 clone and to identify the most easily scored RFLP for mapping purposes. From the 28 markers placed in the *Pto* region, 18 were found to detect an RFLP between the *Pto* NILs with at least one enzyme (64%). Most informative markers detected RFLPs with one to three enzymes on these surveys, although two
20 markers - TG538 and R53, detected polymorphisms with all enzymes tested. Subsequently, TG538 was found to detect RFLPs with an additional 7 restriction enzymes. Overall, marker representation was: 3 cDNAs, 4 RAPDs, 10 RFLP markers, and 1 known gene (TM5). The number of informative markers
25 identified from each approach outlined above was: genome-wide mapping (9 markers); multiprobing (4); RAPD analysis (4).

Development and screening of populations segregating for Pto

30

The majority of the identified markers cosegregated when placed on population 86T64 -- presumably due to a combination of small population size, lower recombination in this wide cross, and close proximity of the markers. To
35 determine the order of the markers and to estimate linkage distances between them and the *Pto* gene, we developed a population that was segregating for *Pst* resistance conferred by *Pto*.

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An F_2 population of approximately 1200 plants was developed from a cross between NILs. Since *Pto* displays dominant gene action, it is necessary to progeny-test any plants resistant to *Pst* with potential recombination events in the *Pto* region to determine the allelic state at the *Pto* locus. In order to avoid progeny testing a large number of plants, we chose to identify and analyze only those plants that were homozygous recessive (*pto/pto*). To accomplish this, we relied on the unusual observation made by French plant breeders that an organophosphorus insecticide, Fenthion, elicits small necrotic lesions on tomato plants carrying the dominant *Pto* locus (Laterrot, 1985; Laterrot and Moretti, 1989, which are hereby incorporated by reference). It is unknown whether this reaction is a pleiotropic effect of the *Pto* gene or the result of a tightly linked gene, termed *Fen*. Whatever the case may be, no plant showing recombination between insensitivity to Fenthion and susceptibility to *Pst* has been identified in populations of over 650 plants, making this an ideal screen for identifying homozygous susceptible plants.

Approximately 1200 F_2 plants were treated with Fenthion, and only those healthy plants (251 total) showing insensitivity (no necrotic lesions) were selected for follow-up. Subsequent work showed that 82% of the plants initially scored as insensitive to Fenthion were susceptible to *Pst* (*pto/pto*). Another 16% were heterozygous at *Pto* and 2% were homozygous resistant (*Pto/Pto*). A second screen of the 18% misscored plants found that they were in fact sensitive to Fenthion. Thus, pre-screening with the insecticide was not absolutely predictive of the *Pto* allelic state but did greatly reduce the amount of progeny testing required. A subsequent screening of a segregating population of 419 plants with Fenthion, where the treatment was modified by dipping the plants in a solution of Fenthion instead of spraying them, resulted in a 97% accurate prediction of *pto/pto* plants (14 misscored plants). Of the 14 misscored plants that were *Pst* resistant all exhibited Fenthion sensitivity when rescreened.

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High resolution linkage analysis

The 251 selected plants were transplanted in the field and analyzed with flanking markers CD31 and TG619 to detect recombinants in the *Pto* region. A total of 85 such plants were identified and these were then analyzed with the remaining 16 informative markers described above. The 18 markers mapped to 9 loci and spanned a region of almost 20 cM. This is shown in Figure 1 which is a linkage map of a 20cM region of tomato chromosome 5 developed from the F_2 population segregating for *Pto*. Notably, crossover events identified between many markers that cosegregated in the 86T64 population and the NILs map (population 90GM251) display almost a 10-fold expansion in the *Pto* region. Over one-half of the map expansion can be accounted for by the distance between TG504 and TG538 (12 cM). In contrast to the TG504-TG538 interval, elsewhere 13 of the markers were found to cluster in a 0.6 cM region. The linkage analysis also revealed that one marker, TG538, cosegregated with the *Pto* locus.

Considering the size of the population and the corresponding standard error, TG538 lies less than 0.6 cM (95% confidence interval) from *Pto*. This discovery of tight linkage to *Pto* is especially interesting in light of the fact that TG538 is clearly derived from a region highly divergent between *L. esculentum* and *L. pimpinellifolium*, as evidenced by the high level of polymorphism detected by this marker (See Figure 1).

Determination of physical distance in the Pto region

Because our goal is to use the linkage map to isolate the *Pto* gene, we used pulsed field gel electrophoresis (PFGE) to estimate the maximum physical distance encompassed by the intervals on each side of TG538. A total of 8 rare-cutting restriction enzymes were surveyed (*Bss*HI, *Nar*I *Nru*I, *Mlu*I, *Sac*II, *Sfi*I and *Sma*I). Those five enzymes which gave fragments between 100 and 900 kb when probed with TG538 were followed up by probing with TG475 and TG504.

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The experiments revealed that TG538 and TG475 detected 19 identical restriction fragments ranging in size from 340 kb to more than 800 kb. In only two enzyme digests, *MluI* and *SfiI*, were unique fragments identified to
5 differentiate these two markers. In all cases, the two NILs were distinguished by RFLPs using PFGE. For TG475, the degree of polymorphism was even higher than when six-base-pair recognition enzymes were used. Although the smallest fragment hybridizing to both TG538 and TG475 was 340 kb (with *NruI*),
10 this fragment only occurred in the *Pst* susceptible line, Rio Grande. Since insertions or deletions could exist in this region that differ between the resistant and susceptible lines, we were primarily interested in the smallest common fragment that existed in Rio Grande-PtoR. The analysis showed
15 that two fragments of 435 and 450 kb were in common between TG475 and TG538 (*SalI* and *SfiI* digests) in Rio Grande-PtoR. Thus, TG475 and TG538 are located no further apart than this distance on the chromosome. The minimum distance between them cannot be determined from these data. A distance of only 400-
20 500 kilobases indicated that we would be able to use chromosome walking (map-based cloning) to isolate the Pto gene.

25 B. Using TG538 to isolate a YAC clone spanning Pto region

RFLP marker TG538 was used to screen a tomato YAC library and a 400 kilobase (kb) clone, PTY538-1, was identified as hybridizing to this marker. End-specific probes corresponding to the right (PTY538-1R) and left (PTY538-1L)
30 arms of PTY538-1 were isolated by inverse PCR and placed on the high resolution linkage map of the region. PTY538-1L mapped 1.8 centimorgans from *Pto*, while PTY538-1R cosegregated with *Pto*. In order to confirm that PTY538-1R encompassed *Pto*, it was necessary to identify a plant with a recombination
35 event between PTY538-1R and *Pto*. We therefore used markers TG538 and PTY538-1R to analyze a total of 1300 plants from various F_2 populations, F_3 families, and over 50 cultivars. One plant was homozygous for the PTY538-1R allele associated

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with *pto* (susceptible allele). All progeny from this plant were resistant to *Pst*, indicating that the plant was homozygous *Pto/Pto*. This result indicated that PTY538-1 spanned the *Pto* locus. Figure 2A shows an example of screening the YAC library with DNA markers. Figure 2B shows a gel separation of a YAC using pulsed field gels. PTY538-1 is at 400kb. Another YAC (not relevant) was analyzed at 600kb. Figure 3 shows genetic mapping of two end clones from PTY538-1.

C. Screening a leaf tissue cDNA library with PTY538-1.

DNA from PTY538-1 was isolated from agarose after separation on a clamped homogeneous electric field (CHEF) gel, and used to probe approximately 920,000 plaque-forming units of a leaf cDNA library.

The cDNA library was constructed by inoculating (i.e. dipping) six-week-old plants of Rio Grande-PtoR and TA208 (*Pto/Pto*) into a solution of avirulent *Pst* strain PT11 (4×10^7 colony-forming-units/ml), 10 mM $MgCl_2$, and 0.05% L-77 Silwet (Union Carbide, Southbury, CT) dispersed in distilled water. Leaf tissue was harvested at 2, 6, 22, 48, and 72 hours after inoculation, polyA⁺ RNA was prepared from each sample, and equal amounts were pooled before library construction. The cDNA library was constructed in vector λ gt10 using a mixture of random and oligo(dT) primers (Stratagene, La Jolla, CA).

From approximately 200 hybridizing plaques, 30 were investigated further. The cDNA inserts were amplified by PCR and used to probe a tomato mapping population consisting of 85 plants with recombination events in the *Pto* region. Two of the clones, CD127 and CD146 (both 1.2 kb), contained sequences that cross-hybridized. When CD127 was mapped, it cosegregated with *Pto*, as shown in Figure 4. The genetic cosegregation of CD127 with *Pto* and the fact that the cDNA was isolated from a leaf tissue library made this cDNA a strong candidate for the *Pto* gene.

CD127 hybridized to numerous polymorphic fragments when probed on blots of genomic DNA from Rio Grande-PtoR and

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Rio Grande plants. This indicated that the clone might contain exons spanning a large region or that it represented a family of related genes. To distinguish between these possibilities, we probed the leaf cDNA library with the CD127 insert and isolated an additional 14 cross-hybridizing clones, ranging from 0.6 to 2.4 kb. The cDNA clone CD186 was isolated from this rescreening of the cDNA library.

Oligonucleotide primers were designed using partial sequence data from both ends of CD127, and used in PCR to amplify a product from the insert of the cDNA clones. PCR products were digested with restriction enzymes recognizing 4-base pair sites (e.g., HaeIII, HinfI, TaqI), and the fragments were separated in a gel composed of 3% Nusieve GTG agarose (FMC) and 1% ultrapure agarose. Six different cDNA types were identified in Rio Grande-PtoR based on their pattern of restriction fragments with homology to CD127. To investigate the genome location of the family members, total DNA from the YAC transformant PTY538-1 was digested with BstNI and analyzed by DNA blot hybridization. The YAC contained all of the CD127-hybridizing fragments, with the exception of a 5 kb band that is common to both Rio Grande-PtoR and Rio Grande. CD127, therefore, represents a gene family that is clustered primarily at the Pto locus. This is shown in Figure 5.

Example 2 - Determination of the DNA sequence of CD127 and CD186.

A. Transfer of cDNA inserts to vectors for DNA sequencing.

CD127 and CD186 were originally isolated as cDNA clones in the cloning vector lambda gt10 (see Figure 6). Because this vector is difficult to manipulate we re-cloned the inserts from CD127 and CD186 into a new vector pcDNAII (see Figure 7), from Invitrogen, Corp., San Diego, California. The inserts were removed from lambda gt10 as EcoRI fragments and cloned into the EcoRI site of pcDNAII, creating two new plasmids: PTC1 (CD127 in pcDNAII) and PTC3 (CD186 in pcDNAII). See Figure 8 which is a map of plasmid vector PTC3.

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These plasmids were used for sequencing the cDNA inserts using standard dideoxy double stranded sequencing techniques (Sequenase Kit, United States Biochemical Corp., Cleveland, OH) and oligonucleotide primers designed to prime at approximately 200 base pair intervals throughout the sequence. Some DNA sequence was also determined at the Purdue University Center for AIDS Research using an automated DuPont Genesis 2000 Instrument. The entire sequence was determined on both strands. The resulting sequence data was analyzed using the program MacVector and overlapping fragments were aligned to create one contig spanning the entire insert.

B. Determination of Sequences.

The entire 2443 base pair DNA sequence of the CD186 insert (SEQ. ID. No. 3) was determined to be as follows:

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1 GAATTCGGCA CGAGCTTAAA TAATGTTATT TGAAGGTTAT TAAGTTGTAC TCAAGTCTCA
61 ATCATGGGAA GCAAGTATTC TAAGGCAACA AATCCATAA ATGATGCTTT AAGCTCGAGT
121 TATCTCGTTC CTTTGTAAAG TTATCGAGTT CCTTTAGTAG ATTTGGAGGA AGCAACTAAT
181 AATTTTGTAT ACAAGTTTTT AATTGGACAT GGTGTCTTTG GGAAGGTTTA CAAGGGTGTT
241 TTGCGTGATG GAGCAAAGGT GGCCCTGAAA AGGCGTACAC CTGAGTCCTC ACAAGGTATT
301 GAAGAGTTTC AAACAGAAAT TGAGACTCTC TCATTTTGCA GACATCCGCA TCTGGTTTCA
361 TTGATAGGAT TCTGTGATGA AAGAAATGAG ATGATTCTAA TTTATAAATA CATGGAGAAT
421 GGGAACCTCA AGAGACATTT GTATGGATCA GATCTACCCA CAATGAGCAT GAGCTGGGAG
481 CAGAGGCTGG AGATATGCAT AGGGGCAGCC AGAGGTCTAC ACTACCTTCA TACTAGAGCA
541 ATTATACATC GTGATGTCAA GTCTATAAAC ATATTGCTTG ATGAGAATTT TGTGCCAAAA
601 ATTACTGATT TTGGAATATC CAAGAAAGGG ACTGAGCTTG ATCAAACCCA TCTTAGCACA
661 GTAGTGAAAG GAACTCTCGG CTACATTGAC CCTGAATATT TTATAAAGGG ACGACTCACT
721 GAAAAATCTG ATGTTTATTC TTTCGGTGTT GTTTTATTCG AAGTTCCTTG TGCTAGGTCT
781 GCCATAGTTC AATCTCTTCC AAGGGAGATG GTTAATTTAG CTGAATGGGC AGTGGAGTCC
841 CATAATAATG GACAGTTGGA ACAAATCGTA GATCCCAATC TTGCAGATAA AATAAGACCA
901 GAGTCCCTCA GGAAGTTTGG AGATACAGCG GTAAATGCT TAGCTTTGTC TAGTGAAGAT
961 AGGCCATCAA TGGGTGATGT GTTGTGGAAA CTGGAGTATG CACTTCGTCT CCAAGAGTCT
1021 GTTATTTAAG ATATTTTGTG TTTTCTGAGT TTTATATAGA AAAAGGTAAA CTTTGAAGAAC
1081 TTGAATTGCT ATACCTGTGG ATCCTTCTTT CATTTTATTA GGTGCGTCCG GCTGTTACAC
1141 ATATTGTATA TGGTTCTTAT TAAGTTCTTC AGACATTTTG TTATTGTAAA GAGGCAAAAA
1201 GGAAGTTTGC TGCTTTGACA TAGTCAATCT AAAACTATAT ACATTCAACT TTCAGAATGG
1261 AACTATAAAA GTTTGTGGAG CAATTCAAAA TGTACTCAA CCTGTTTACA AAATGACTAT
1321 TGTAGAGCAA TAATGGTTAT AATATATAAC CATTATTGAG TAATATTTTT GTAGTAGTAT
1381 TGCCCAAGTC CATTAGCGGA GAGGTAATTT TCTTTTGGT TCTCTCTTCC ACAATAGCTA
1441 TCAATCTCTC TGTCTTCTCG CTAAATTTCC TCAGTTGTGG TATAATCAGA GGTTCTTAAG
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1501 CCTTCTGTTT TGTATACATA TATTGTGAT TTTCATCTAT CATGCTTACT GTTAGGAGTT
1561 ATATTGCTTG ATGAGAATTT TGTGGCAAAA ATTAATGATT TTGGTCGATT CAAGAAGCTT
1621 GATCAAACCC ATGTTACCAC AATAGTAAAG GAACTTTGG TTACCTTGAC CCTGAATATT
1681 ATCAAACTAG TCAGCTGACA GAAAATCTG ATGTTTATTC TTTCGGTGGT GTTTTATTAG
1741 AAGTTATTTG TGCTAGGCCT GCGCTGGATT CATCTCGTTC GAGGGAGATG GTCAGCTCAG
1801 TTAAATGGGC AAAGGAGTGT CAGAAGAACG GACAGTCGGA ACGAATTATA GATCCCAATC
1861 TTGTTGCGAA AATAAGACCA GATTCCCCCA GGAAGTTTGG AGAAACAGCT GTGAAATGCT
1921 TAGCTGAAAC TGGCGTAAAC AGGCCATCAA TGGGTGAGGT GCTCGAGAAA CTGGACTATG
1981 CACTTCATCT CTAAGAGCCT GTCATTCAAG AAAACAGTAC CATCCCTATC CGCGAGCAAA
2041 TCAATGATTT CAGTCATGTT GATGACACTT CCTCTGCTTC TTCGGTCAAA ATTGGGCTGA
2101 TCTCTAGTAT GAATGCGTTC AGATTTTGCT CAAGAAAACA GCCGGGAGAA GTTCAATTAA
2161 TGGTTGCACT CCATGGGAAC CAACTATTCC AAGCCAACAA CTTCCATAAA TGATGCTTCC
2221 AATTTGAGTA ATCGCGTTCC TTTTGAAAGT TTTCGAGTTC CTTTGTAGA TTGTCAGGAA
2281 GCAACTAATA ACTTTGATGA CAAGTTTCTG ATTGGAGTGG GTATATTTGG TAAGGATTAC
2341 AGGGGTGTTT TCGGTGATGG TACAAAGGTG GCCCTGAAAA GACATAAGCC TGAGTCTCCA
2401 CAAGGTATTG AAGAGTTCCG AACAGAAATC TCGTACCGAA TTC

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A 963 bp open reading frame sequence (ORF1) (SEQ. ID. Nos. 1 and 2) was found in the region nearest to the 35S CaMV promoter in pPTC8 (see Figure 9). ORF1, hereafter referred to as *Pto*, encodes a 321 amino acid hydrophilic protein. ORF1 and the corresponding amino acid sequence were identified using the program MacVector.

Example 3 - Incorporation of CD127 and CD186 inserts into plant expression vectors.

A. Cloning of CD127 and CD186 inserts into expression vector pBI121.

The cloning of CD127 and CD186 inserts into expression vectors began with the plasmids PTC1 and PTC3. To prepare the inserts of PTC1 and PTC3 for transformation into plants, the inserts were first cloned into the Ti-based plant transformation vector pBI121 (see Figure 10) from Clontech Laboratories, Palo Alto, California. Two cDNA clones, representative of the two size classes of transcripts, were subcloned into pBI121: (CD127 [1.2 kb] and CD186 [2.4kb]). Based on the DNA sequence information, the sense orientation of each fragment was determined and the cDNA inserts were placed in the sense orientations in pBI121 under the

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transcriptional control of the constitutive cauliflower mosaic virus (CaMV) 35S promoter. The resulting plasmids were designated pPTC5 (CD127, [PTC1 insert]) and pPTC8 (CD186, [PTC3 insert]) (see Figure 9). The constructs were introduced into *Agrobacterium tumefaciens* LBA4404 by electroporation. The bacteria containing pPTC5 and pPTC8 were used to transform cotyledon explants of *Pst*-susceptible tomato cultivar, Moneymaker.

10 Example 4 - Method for Transforming Tomato

Sterile cotyledon explants of Example 3 from approximately week old seedlings were incubated for 10 minutes in an *Agrobacterium tumefaciens* (LBA4404) harboring either the CD186 or CD127 T-DNA plasmid. Explants were then transferred to TRS medium (containing 70 mg/l kanamycin as a selective agent for plant cells harboring the T-DNA insertion) with media changes as needed at approximately 1 month periods or until shoots had formed.

20 The TRS medium was prepared by blending 500 ml of water with 100 ml MS major salts, 10 ml MS minor salts, 5 ml MS iron stock, 1 ml B5 vitamin stock, and 30 g sucrose. The pH was measured to ensure that it was at 5.8. 500 ml of the mixture was blended with 4.0 g agar, autoclaved, and cooled to 45°C. To the cooled mixture, 0.5 ml zeatin (1 mg/ml), 0.05 ml IAA (1 mg/ml), 1 ml kanamycin (25 mg/ml), and 1 ml timentin (200 mg/ml) or 1.25 ml carbenicillin (200 mg/ml) were added.

5 mm shoots were excised and placed on P rooting medium. P rooting medium was prepared by blending 100 ml P major salts, 1 ml P minor salts, 5 ml MS iron stock, 2.5 ml MS thiamine stock, 10 ml MS myoinositol stock, 1 ml nicotinic acid (0.5 mg/ml), 1 ml pyridoxine (0.5 mg/ml), 1 ml glycine (2 mg/ml), and 30 g sucrose and the volume of that mixture was adjusted with distilled water to 500ml. The pH of the mixture was then measured to ensure that it had a pH of 5.8, and 4.0 g Agar was added to 500 ml of the mixture. The mixture was then autoclaved and cooled to 45°C. 0.09 ml IAA (1 mg/ml), 1.0 ml

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kanamycin (25 mg/ml), and 1 ml timentin (200 mg/ml) were added to the mixture.

Healthy green plants with good root formation were then removed from magenta boxes, transferred to a soil mix, moved to a greenhouse, and screened with *Pseudomonas syringae*.

Putative transformed plants were verified through analysis of genomic plant DNA using both a PCR assay for presence of the CaMV 35S promoter (forward primer = AAAGGAAGGTGGCTCCTACAAAT (SEQ ID. No. 4), reverse primer = CCTCTCCAAATGAAATGAACTTCC (SEQ ID. No. 5)) and via Southern probing using the CaMV35S promoter sequence and CD127 insert DNA as probes. Only plants confirmed to be transformed by one or both of these assays were utilized for further experiments.

Example 5 - Determination of *Pst* resistance in transformed plants.

A. Initial testing for *Pst* resistance in transformed plants.

Five weeks after the transformed plants were transferred to soil, single leaves were inoculated with *Pst* strain T1(pPTE6) carrying *avrPto*. Leaflets were inoculated by dipping into a solution of avirulent *Pst* strain T1(pPTE6) (2×10^7 colony-forming-units/ml), 10 mM $MgCl_2$, and 0.05% L-77 Silwet (Union Carbide, Southbury, CT) dispersed in distilled water. Under these conditions, symptoms of bacterial speck appeared after 5-7 days on susceptible plants. A resistant reaction was indicated by the absence of necrotic specks on the inoculated leaves. A susceptible reaction was indicated by numerous necrotic specks surrounded by chlorotic halos. Reactions were scored 8-10 days after inoculation. As controls, 4 week-old seedlings of Rio Grande-PtoR, Moneymaker, and Moneymaker transformed with pBI121 alone were also inoculated. Of two plants that were confirmed to contain the integrated transgene from pPTC8 (PTC8/39 and PTC8/56) both were resistant to *Pst* strain T1(pPTE6). None of the nine transformants containing integrated copies of pPTC5 displayed resistant phenotypes.

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B. Genetic analysis to confirm that *Pst* resistance is conferred by CD186 transgene.

To confirm that *Pst* resistance was due to
5 introduction of the CD186 cDNA insert, a resistant RO
transgenic plant (PTC8/39) was crossed to a susceptible
control plant (cv. Rio Grande). Of 22 backcross progeny
examined, 9 inherited the CD186 transgene. This closely fit a
1:1 segregation ratio and indicated that the original
10 integration of pPTC8 sequences in PTC8/39 occurred at a single
locus. The same 9 plants containing the CD186 transgene
displayed no disease symptoms upon inoculation with T1(pPTE6).
This indicates that CD186 transgene was sufficient for
conferring resistance in a normally *Pst*-susceptible tomato
15 cultivar. The remaining 13 plants lacked the CD186 transgene
and displayed typical symptoms of bacterial speck. All 22
progeny plants were susceptible to *Pst* strain T1 which lacks
avrPto. Because it is possible that plants showing no disease
symptoms in response to T1(pPTE6) might still harbor a large
20 population of *Pst*, we monitored the colony-forming-units of
Pst in the progeny plants and in control over plants over a
period of 4 days after inoculation and plotted the number of
leaf bacteria versus time (see Figure 11). Figure 10 shows
the growth of *Pst* in the leaves of 7-week-old Rio Grande-PtoR
25 ("RG-R"), Moneymaker ("MM"), backcross progeny with pPTC8
("BC-R"), and without pPTC8 ("BC-S") lines of tomato which
were inoculated with *Pst* strain T1(pPTE6) and then had
bacterial populations determined at specified points in time.
The plotted values were means of 3 examples, each consisting
30 of 3 leaf disks. Error bars indicate standard deviations.
The 9 progeny (BC-R) exhibiting no disease symptoms contained
10³-fold fewer bacteria per cm² leaf area than the BC-S
susceptible plants at the end of this time period. Lower
bacterial populations in BC-R plants than in Rio Grande-PtoR
35 plants were observed and may be the result of a higher
abundance of *Pto* protein in BC-R plants due to the
constitutive 35S promoter. Thus, CD186 functionally
complements *Pto* in *Pst*-susceptible plants by inhibiting growth

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of the *Pst* population and suppressing symptoms of bacterial speck disease.

Example 6 - Determining if DNA sequences in other crop species
5 have homology to *Pto*.

A. Southern blot analysis of genomic DNA from a variety of
crop species using as a probe CD127 and CD186 insert DNA.

10 To determine if homologs of the CD127 gene family
were present in other plant species, we performed Southern
blot analysis on genomic DNA isolated from the following
plants and digested with *Eco*R1 (amount loaded on gel is
indicated): tomato (Rio Grande-*Pto*R, 3 ug); pepper (*Capsicum*
15 *annum*, 5 ug); petunia (*Petunia parodii*, 5 ug); tobacco
(Samsun, 5 ug); *Arabidopsis* (Col-0, 1 ug); bean (*Phaseolus*
acutifolius, G40178, 3 ug); bean (*P. acutifolius*, PO310800, 3
ug); soybean (Centennial, 4 ug); pea (Sparkle, 10 ug); rice
(IRAT, 3 ug); maize (RI 24, 15 ug); barley (SE16, 15 us);
20 wheat (R-4, 15 ug); sugarcane (*Saccharum spontaneum*, SES208,
10 ug). The digested genomic DNA was separated by
electrophoresis on a 1% agarose gel. The gel was blotted onto
Hybond N+ membrane which was hybridized with radiolabeled
CD127 insert using random-hexamer ³²P-labeled (a.p. Feinberg,
25 B. Bogelstein, Anal. Biochem. 132, 6 [1938]) PCR product (1-2
X 10⁶ cpm/ml buffer) amplified from the cDNA clone. Filter was
washed to 0.5X SSC at 65°C and exposed to film for 24 hours
for the Solanaceous species (lanes 1-6) and 7 days for the
remaining species (lanes 7-16). Figure 12 shows the resulting
30 DNA blot analysis of the species distribution of *Pto* gene
homologs. Homologs of CD127 were identified in all species
examined. Multiple bands were detected in many of the
species, indicating the possible presence of a gene family
similar to that in tomato. This sequence conservation
35 indicates that other plant species appear to contain genes
with structural, and perhaps functional, similarity to the
CD127 gene family. Consistent with this, some soybean

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cultivars exhibit hypersensitive resistance in response to *avrPto*.

Similar experimentation using CD186 as a probe yielded results substantially identical to that achieved with CD127.

Example 7 - Homology of CD186 with Other Known Genes

Figure 13A is a physical map of the CD186 cDNA insert for ORF1 (*Pto*) and the 35 S CaMV promotor on pPTC8 with the regions designated a' and b' being homologous to a and b on ORF1 and maybe representing a downstream pseudogene. The numbers above the map are DNA base pairs, with the numbers in parentheses referring to positions in ORF1 corresponding to a' and b'. Figure 13B is the deduced amino acid sequence of ORF1 (*Pto*) shown in standard 1-letter code.

The deduced amino acid sequences from the open reading frames of CD186 and CD127 were run against Genbank release 77 using the BLAST program (Altschul, et al. J. Mol Biol 215:403 (1990)). Highly significant matches were found with a variety of protein kinases from plants and animals. Eleven subdomains, including 15 invariant amino acids, characteristics of protein kinases, were also found to be present in both the CD186 and CD127 amino acid sequences. Furthermore, sequences indicative of serine/threonine kinases occur in subdomains VI (consensus DLKPEN) and VIII (consensus (G(T/S)XX(Y/F)XAPE). This is shown in Figure 13B where the positions of these subdomains characteristic of protein kinases are indicated in parenthesis above the sequence, while the amino acids that are highly conserved among protein kinases are underlined. Residues that indicate serine/threonine specificity are double-underlined. A site at the N-terminus is overlined which may be modified by the addition of myristic acid (a potential myristoylation site). To examine further the relationship to protein kinases, the CD186 amino acid sequence was compared to the Protein Kinase Catalytic Domain Database (Hanks and Quinn Meth Enzymol 200:38 (1991)) using the align function of MacVector sequence

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analysis software (Kodak International Biotechnologies, version 4.0.1). The five most similar matches were to putative serine/threonine protein kinase genes of plant origin (L00670, M84659, ZMPK1), including three with unknown roles from *Arabidopsis* (TMK1, GenBank accession number L00670; ARK1, M80238; RLPK, M84659) one with an unknown role from maize (ZMPK1, X52384), and one from *Brassica* (SRK6, M76647) that is believed to be involved in pollen/stigma recognition and, like tomato-*Pst* to be based on a gene for gene interaction (pollen/cell/stigma) (SRK6, M76647) (Stein et al. Proc Natl Acad Sci USA 88:8816 (1991)). The similarity between *Pto* and SRK6 is particularly interesting since SRK6 appears to be involved in a specific cell-cell interaction (pollen cell/stigma papillar cell) that, like tomato-*Pst*, is based on a gene-for-gene relationship. Other than plants, the closest matches in the data base were to mammalian serine/threonine kinases of the *Raf* family (Bonner et al. Nucl Acids Res 14:1009 (1986) and (MacIntyre et al. Mol Cell Biol 7:2135 (1987)).

Example 8 - Examination of differences in transcript size or abundance between tomato lines for resistance *Pst*.

A. Analysis by Northern blots of transcript size and abundance.

RNA blot analysis (Figure 14) was used to determine if there were differences in transcript size or abundance produced by the CD127 family members among tomato lines resistant or susceptible to *Pst*. PolyA+RNA of Rio Grande-*PtoR* (*Pto/Pto*), Spectrum 151 (*Pto/pto*); and Moneymaker (*pto/pto*) was isolated from leaf tissue of 5 week-old plants, separated on a 1.4% agarose-formaldehyde gel and blotted into nitrocellulose. The blot was hybridized with ³²P-labeled CD127 insert. A difference in abundance among the samples was mostly attributable to unequal loading of RNA as indicated by hybridizing the identical filter with a probe for ribulose biphosphate carboxylase (Rubisco) transcript. (The Rubisco

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probe detects a gene transcript that is expected to be expressed equally in all tomato lines examined). A prominent 1.3 kb band was observed in resistant and susceptible lines. A fainter band of 2.5 kb may indicate the presence of less abundant, longer transcripts in the CD127 family. No obvious induction of gene expression upon infection with *Pst* was observed. Thus, we could find no difference in either abundance or transcript size and the basis of CD127 action is probably not due to a transcript of difference size being produced by the resistant line.

Although the invention has been described in detail for the purpose of illustration, it is understood that such detail is solely for that purpose, and variations can be made therein by those skilled in the art without departing from the spirit and scope of the invention which is defined by the following claims.

- 32 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Tanksley, Steven D.
Martin, Gregory B.

(ii) TITLE OF INVENTION: GENE CONFERRING DISEASE
RESISTANCE TO PLANTS BY RESPONDING TO AN AVIRULENCE GENE IN
PLANT PATHOGENS

(iii) NUMBER OF SEQUENCES: 5

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Michael L. Goldman
(B) STREET: Clinton Square, P.O. Box 1051
(C) CITY: Rochester
(D) STATE: New York
(E) COUNTRY: U.S.A.
(F) ZIP: 14603

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Goldman Mr., Michael L.

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (716)263-1000
(B) TELEFAX: (716)-263-1600

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 321 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Met	Gly	Ser	Lys	Tyr	Ser	Lys	Ala	Thr	Asn	Ser	Ile	Asn	Asp	Ala	Leu	1	5	10	15
Ser	Ser	Ser	Tyr	Leu	Val	Pro	Phe	Glu	Ser	Tyr	Arg	Val	Pro	Leu	Val	20	25	30	
Asp	Leu	Glu	Glu	Ala	Thr	Asn	Asn	Phe	Asp	His	Lys	Phe	Leu	Ile	Gly	35	40	45	
His	Gly	Val	Phe	Gly	Lys	Val	Tyr	Lys	Gly	Val	Leu	Arg	Asp	Gly	Ala	50	55	60	
Lys	Val	Ala	Leu	Lys	Arg	Arg	Thr	Pro	Glu	Ser	Ser	Gln	Gly	Ile	Glu	65	70	75	80
Glu	Phe	Glu	Thr	Glu	Ile	Glu	Thr	Leu	Ser	Phe	Cys	Arg	His	Pro	His	85	90	95	
Leu	Val	Ser	Leu	Ile	Gly	Phe	Cys	Asp	Glu	Arg	Asn	Glu	Met	Ile	Leu	100	105	110	
Ile	Tyr	Lys	Tyr	Met	Glu	Asn	Gly	Asn	Leu	Lys	Arg	His	Leu	Tyr	Gly	115	120	125	
Ser	Asp	Leu	Pro	Thr	Met	Ser	Met	Ser	Trp	Glu	Gln	Arg	Leu	Glu	Ile	130	135	140	
Cys	Ile	Gly	Ala	Ala	Arg	Gly	Leu	His	Tyr	Leu	His	Thr	Arg	Ala	Ile	145	150	155	160
Ile	His	Arg	Asp	Val	Lys	Ser	Ile	Asn	Ile	Leu	Leu	Asp	Glu	Asn	Phe	165	170	175	
Val	Pro	Lys	Ile	Thr	Asp	Phe	Gly	Ile	Ser	Lys	Lys	Gly	Thr	Glu	Leu	180	185	190	
Asp	Gln	Thr	His	Leu	Ser	Thr	Val	Val	Lys	Gly	Thr	Leu	Gly	Tyr	Ile	195	200	205	
Asp	Pro	Glu	Tyr	Phe	Ile	Lys	Gly	Arg	Leu	Thr	Glu	Lys	Ser	Asp	Val	210	215	220	
Tyr	Ser	Phe	Gly	Val	Val	Leu	Phe	Glu	Val	Leu	Cys	Ala	Arg	Ser	Ala	225	230	235	240
Ile	Val	Gln	Ser	Leu	Pro	Arg	Glu	Met	Val	Asn	Leu	Ala	Glu	Trp	Ala	245	250	255	

Val Glu Ser His Asn Asn Gly Gln Leu Glu Gln Ile Val Asp Pro Asn
 260 265 270

Leu Ala Asp Lys Ile Arg Pro Glu Ser Leu Arg Lys Phe Gly Asp Thr
 275 280 285

Ala Val Lys Cys Leu Ala Leu Ser Ser Glu Asp Arg Pro Ser Met Gly
 290 295 300

Asp Val Leu Trp Lys Leu Glu Tyr Ala Leu Arg Leu Gln Glu Ser Val
305 310 315 320

Ile

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 966 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

SUBSTITUTE SHEET (RULE 26)

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ACTGATTTTG GAATATCCAA GAAAGGGACT GAGCTTGATC AAACCCATCT TAGCACAGTA 600
 GTGAAAGGAA CTCTCGGCTA CATTGACCTT GAATATTTTA TAAAGGGACG ACTCACTGAA 660
 AAATCTGATG TTTATTCTTT CGGTGTTGTT TTATTGAAG TTCTTTGTGC TAGGTCTGCC 720
 ATAGTTCAAT CTCTTCCAAG GGAGATGGTT AATTAGCTG AATGGGCAGT GGAGTCGCAT 780
 AATAATGGAC AGTTGGAACA AATCGTAGAT CCAATCTTG CAGATAAAAT AAGACCAGAG 840
 TCCCTCAGGA AGTTTGGAGA TACAGCGGTA AAATGCTTAG CTTTGTCTAG TGAAGATAGG 900
 CCATCAATGG GTGATGTGTT GTGGAACTG GAGTATGCAC TTCGTCTCCA AGAGTCTGTT 960
 ATTTAA 966

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2443 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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 TATCTCGTTC CTTTGGAAAG TTATCGAGTT CCTTTAGTAG ATTTGGAGGA AGCAACTAAT 180
 AATTTTGATC ACAAGTTTTT AATTGGACAT GGTGTCTTTG GGAAGGTTTA CAAGGGTGTT 240
 TTGCGTGATG GAGCAAAGGT GGCCCTGAAA AGGCGTACAC CTGAGTCCTC ACAAGGTATT 300
 GAAGAGTTCG AAACAGAAAT TGAGACTCTC TCATTTTGCA GACATCCGCA TCTGGTTTCA 360
 TTGATAGGAT TCTGTGATGA AAGAAATGAG ATGATTCTAA TTTATAAATA CATGGAGAAT 420
 GGGAACTCA AGAGACATTT GTATGGATCA GATCTACCCA CAATGAGCAT GAGCTGGGAG 480

- 36 -

CAGAGGCTGG AGATATGCAT AGGGGCAGCC AGAGGTCTAC ACTACCTTCA TACTAGAGCA 540
ATTATACATC CTCATCTCAA CTCATTAAC ATATTGCTTG ATGAGAATTT TGTGCCAAAA 600
ATTACTGATT TTGGAATATC CAAGAAAGGG ACTGAGCTTG ATCAAACCCA TCTTAGCACA 660
GTAGTGAAAG GAACTCTCGG CTACATTGAC CCTGAATATT TTATAAAGGG ACGACTCACT 720
GAAAAATCTG ATGTTTATTC TTTCGGTGTT GTTTTATTCG AAGTTCTTTG TGCTAGGTCT 780
GCCATAGTTC AATCTCTTCC AAGGGAGATG GTTAATTTAG CTGAATGGGC AGTGGAGTCG 840
CATAATAATG GACAGTTGGA ACAAATCGTA GATCCCAATC TTGCAGATAA AATAAGACCA 900
GAGTCCCTCA GGAAGTTTGG AGATACAGCG GTAAAATGCT TAGCTTTGTC TAGTGAAGAT 960
AGGCCATCAA TGGGTGATGT GTTGTGGAAA CTGGAGTATG CACTTCGTCT CCAAGAGTCT 1020
GTTATTTAAG ATATTTTTGT TTTTCTGAGT TTTATATAGA AAAAGGTAAA CTTTGAAAAC 1080
TTGAATTGCT ATACCTGTGG ATCCTTCTTT CATTTTATTA GGTGCGTCCG GCTGTTACAC 1140
ATATTGTATA TGGTTCTTAT TAAGTTCTTC AGACATTTTG TTATTGTAAA GAGGCAAAAA 1200
GGAAGTTTGC TGCTTTGACA TAGTCAATCT AAACTATAT ACATTCAACT TTCAGAATGG 1260
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TGCCCAAGTC CATTAGCGGA GAGGTAATTT TCTTTTGGT TCTCTCTTCC ACAATAGCTA 1440
TCAATCTCTC TGTCTTCTCG CTAAATTTCC TCAGTTGTGG TATAATCAGA GGTTCCTAAG 1500
CCTTCTGTTT TGTATACATA TATTGTGAT TTTCATCTAT CATGCTTACT GTTAGGAGTT 1560
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GATCAAACCC ATGTTACCAC AATAGTAAAG GAACTTTTGG TTACCTTGAC CCTGAATATT 1680
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TTAAATGGGC AAAGGAGTGT CAGAAGAACG GACAGTCGGA ACGAATTATA GATCCCAATC 1860
TTGTTGGCAA AATAAGACCA GATTCCCCCA GGAAGTTTGG AGAAACAGCT GTGAAATGCT 1920

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TAGCTGAAAC TGGCGTAAAC AGGCCATCAA TGGGTGAGGT GCTCGAGAAA CTGGACTATG 1980
 CACTTCATCT CTAAGAGCCT GTCATTCAG AAAACAGTAC CATCCCTATC CGCGAGCAAA 2040
 TCAATGATTT CAGTCATGTT GATGACACTT CCTCTGCTTC TTCGGTCAAA ATTGGGCTGA 2100
 TCTCTAGTAT GAATGCGTTC AGATTTTGCT CAAGAAAACA GCCGGGAGAA GTTCAATTAA 2160
 TGGTTGCACT CCATGGGAAC CAACTATTCC AAGCCAACAA CTTCCATAAA TGATGCTTCC 2220
 AATTTGAGTA ATCGCGTTCC TTTTGAAAGT TTCGAGTTC CTTTGTAGA TTTGCAGGAA 2280
 GCAACTAATA ACTTTGATGA CAAGTTTCTG ATTGGAGTGG GTATATTTGG TAAGGATTAC 2340
 AGGGGTGTTT TGGGTGATGG TACAAAGGTG GCCCTGAAAA GACATAAGCC TGAGTCTCCA 2400
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(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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23

(2) INFORMATION FOR SEQ ID NO:5:

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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CCTCTCCAAA TGAAATGAAC TTCC

24

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WHAT IS CLAIMED:

1. An isolated gene fragment conferring disease resistance to plants by responding to an avirulence gene in plant pathogens, wherein said gene fragment encodes for protein kinase.
5
2. An isolated gene fragment according claim 1, wherein the protein kinase is a serine/threonine kinase.
10
3. An isolated gene fragment according claim 1, wherein said gene fragment encodes for the amino acid sequence corresponding to SEQ. ID. No. 1.
4. An isolated gene fragment according claim 3, wherein said gene fragment comprises the nucleotide sequence corresponding to SEQ. ID. No. 2.
15
5. An isolated gene fragment according to claim 3, wherein said gene fragment comprises the nucleotide sequence corresponding to SEQ. ID. No. 3.
20
6. An isolated gene fragment according claim 1, wherein said gene fragment imparts to tomato resistance to *Pseudomonas syringae*.
25
7. An isolated protein conferring disease resistance to plants, wherein said protein comprises an amino acid sequence for protein kinase.
30
8. An isolated protein according to claim 7, wherein the protein kinase is a serine/threonine kinase.
9. An isolated protein according to claim 7, wherein said protein includes the amino acid sequence corresponding to SEQ. ID. No. 1.
35

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10. An isolated protein according to claim 7,
wherein said protein imparts to tomato resistance to
Pseudomonas syringae.

5 11. An isolated protein according to claim 7,
wherein said protein is recombinant.

10 12. A recombinant DNA expression system comprising
an expression vector into which is inserted a heterologous DNA
conferring disease resistance to plants by responding to an
avirulence gene in plant pathogens, wherein said DNA encodes
for protein kinase.

15 13. A recombinant DNA expression system according
to claim 12, wherein the protein kinase is a serine/threonine
kinase.

20 14. A recombinant DNA expression system according
to claim 12, wherein said heterologous DNA encodes for the
amino acid sequence corresponding to SEQ. ID. No. 1.

25 15. A recombinant DNA expression system according
to claim 12, wherein said heterologous DNA is inserted into
said vector in proper orientation and correct reading frame.

 16. A recombinant DNA expression system according
to claim 15, wherein said heterologous DNA comprises the
nucleotide sequence corresponding to SEQ. ID. No. 2.

30 17. A recombinant DNA expression system according
to claim 15, wherein said heterologous DNA comprises the
nucleotide sequence corresponding to SEQ. ID. No. 3.

35 18. A cell transformed with a heterologous DNA
conferring disease resistance to plants by responding to an
avirulence gene in plant pathogens, wherein said heterologous
DNA encodes for protein kinase.

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19. A cell according to claim 18, wherein the protein kinase is a serine/threonine kinase.

20. A cell according to claim 18, wherein said
5 heterologous DNA encodes for the amino acid sequence corresponding to SEQ. ID. No. 1.

21. A cell according to claim 18, wherein said cell
is selected from the group consisting of plant cells and
10 bacterium.

22. A cell according to claim 21, wherein said cell
is a plant cell selected from the group consisting of
gymnosperm, monocot, and dicot.

15

23. A cell according to claim 22, wherein the host
cell is a crop plant cell selected from the group consisting
of rice, wheat, barley, rye, corn, potato, carrot, sweet
potato, bean, pea, chicory, lettuce, cabbage, cauliflower,
20 broccoli, turnip, radish, spinach, asparagus, onion, garlic,
eggplant, pepper, celery, carrot, squash, pumpkin, zucchini,
cucumber, apple, pear, quince, melon, plum, cherry, peach,
nectarine, apricot, strawberry, grape, raspberry, blackberry,
pineapple, avocado, papaya, mango, banana, soybean, tobacco,
25 tomato, sorghum, and sugarcane.

24. A cell according to claim 21, wherein the cell
is from the genus *Agrobacterium*.

30 25. A cell according to claim 18, wherein said
heterologous DNA is inserted in a recombinant DNA expression
system comprising an expression vector.

26. A transgenic plant transformed with a gene
35 fragment conferring disease resistance to plants by responding
to an avirulence gene in plant pathogens, wherein said gene
fragment encodes for protein kinase.

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27. A transgenic plant according to claim 26,
wherein the protein kinase is a serine/threonine kinase.

28. A transgenic plant according to claim 26,
5 wherein said gene fragment encodes for the amino acid sequence
corresponding to SEQ. ID. No. 1.

29. A transgenic plant according to claim 26,
wherein said plant is selected from the group consisting of
10 gymnosperm, monocot, and dicot.

30. A transgenic plant according to claim 29,
wherein said plant is selected from the group consisting of
rice, wheat, barley, rye, corn, potato, carrot, sweet potato,
15 bean, pea, chicory, lettuce, cabbage, cauliflower, broccoli,
turnip, radish, spinach, asparagus, onion, garlic, eggplant,
pepper, celery, carrot, squash, pumpkin, zucchini, cucumber,
apple, pear, quince, melon, plum, cherry, peach, nectarine,
apricot, strawberry, grape, raspberry, blackberry, pineapple,
20 avocado, papaya, mango, banana, soybean, tobacco, tomato,
sorghum, and sugarcane.

31. A process of conferring disease resistance to
plants comprising:
25 growing plant host cells transformed with a
recombinant DNA expression system comprising an expression
vector into which is inserted a heterologous DNA conferring
disease resistance to plants by responding to an avirulence
gene in plant pathogens, wherein said DNA encodes for protein
30 kinase and
expressing the heterologous DNA in the host
cells to confer disease resistance on the host cells.

32. A process according claim 31, wherein the
35 protein kinase is a serine/threonine kinase.

33. A process according to claim 31, wherein the heterologous DNA encodes for the amino acid sequence corresponding to SEQ. ID. No. 1.

5 34. A process according to claim 31, wherein the
plant host cells are selected from the group consisting of
gymnosperm, monocot, and dicot.

35. A process according to claim 34, wherein said
10 plant host cells are selected from the group consisting of
rice, wheat, barley, rye, corn, potato, carrot, sweet potato,
bean, pea, chicory, lettuce, cabbage, cauliflower, broccoli,
turnip, radish, spinach, asparagus, onion, garlic, eggplant,
pepper, celery, carrot, squash, pumpkin, zucchini, cucumber,
15 apple, pear, quince, melon, plum, cherry, peach, nectarine,
apricot, strawberry, grape, raspberry, blackberry, pineapple,
avocado, papaya, mango, banana, soybean, tobacco, tomato,
sorghum, and sugarcane.

20 36. A process according to claim 31, wherein the plant host cells are transformed by a process comprising:
 contacting the plant host cells with an inoculum of a bacterium, wherein the bacterium is transformed with the recombinant DNA expression system.

25

37. A process according to claim 34, wherein the cell is from the genus *Agrobacterium*.

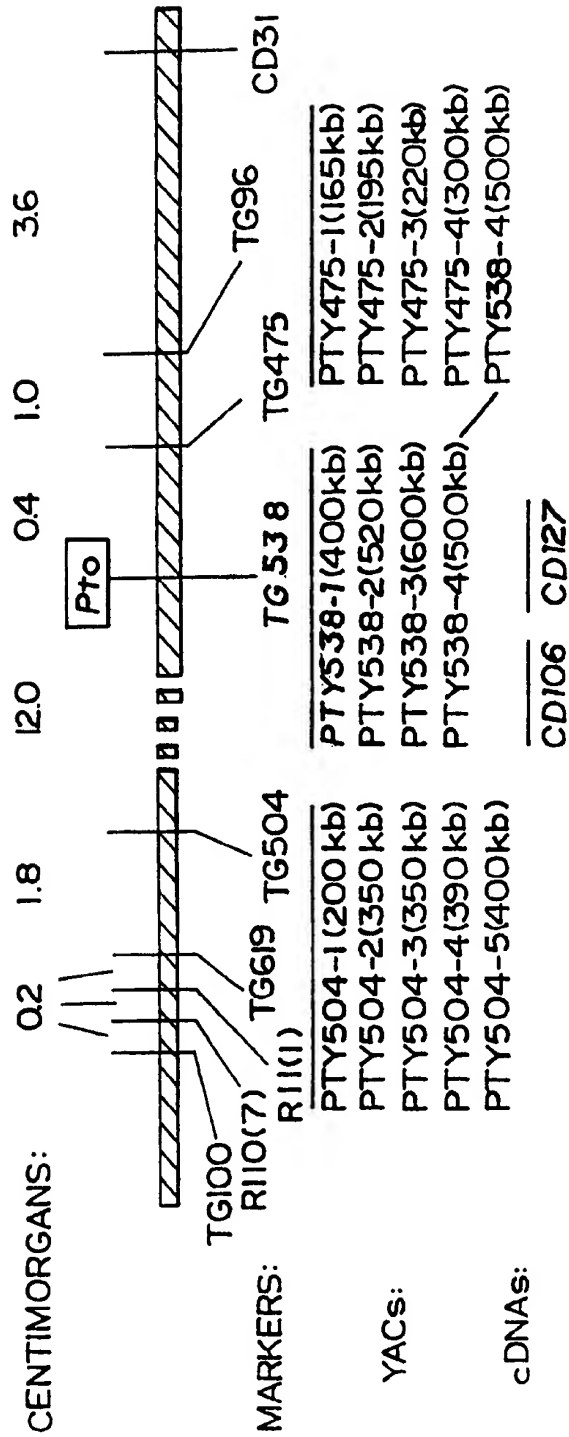
38. A process according to claim 31, wherein the
30 host cells are transformed by a process comprising:
propelling particles at the host cells under
conditions effective for the particles to penetrate into the
cell interior and
introducing the recombinant DNA expression
35 system into the cell interior.

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39. A process according to claim 38, wherein the recombinant DNA expression system is carried into the cell interior together with the particles.

- 5 40. A process according to claim 38, wherein the recombinant DNA expression system surrounds the host cells and is drawn into the cell interior by the particle's wake.

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FIG.1

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Screening tomato YAC library with TG538

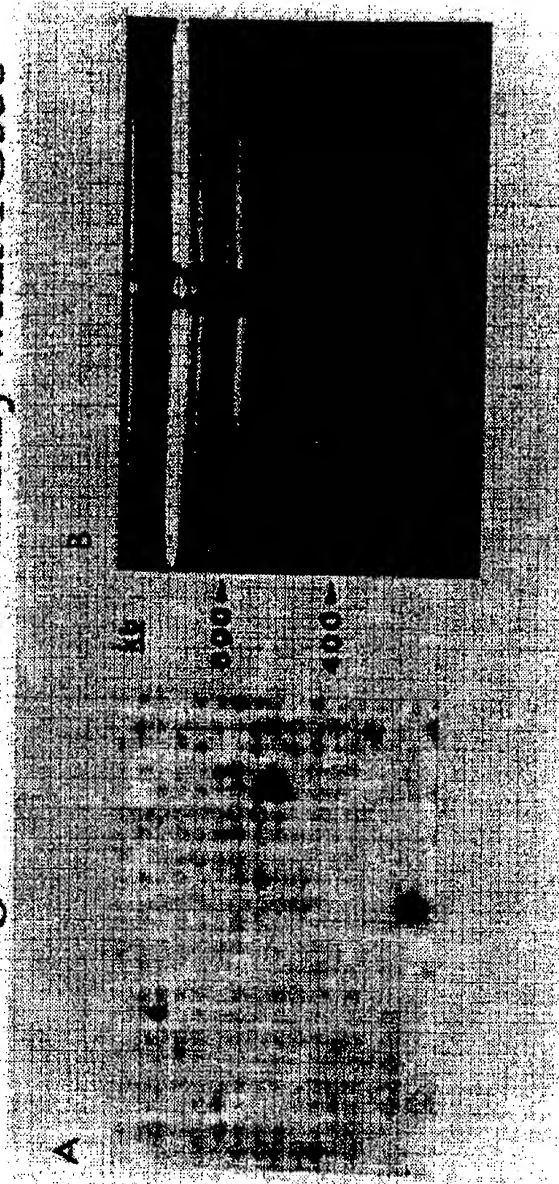


FIG. 2

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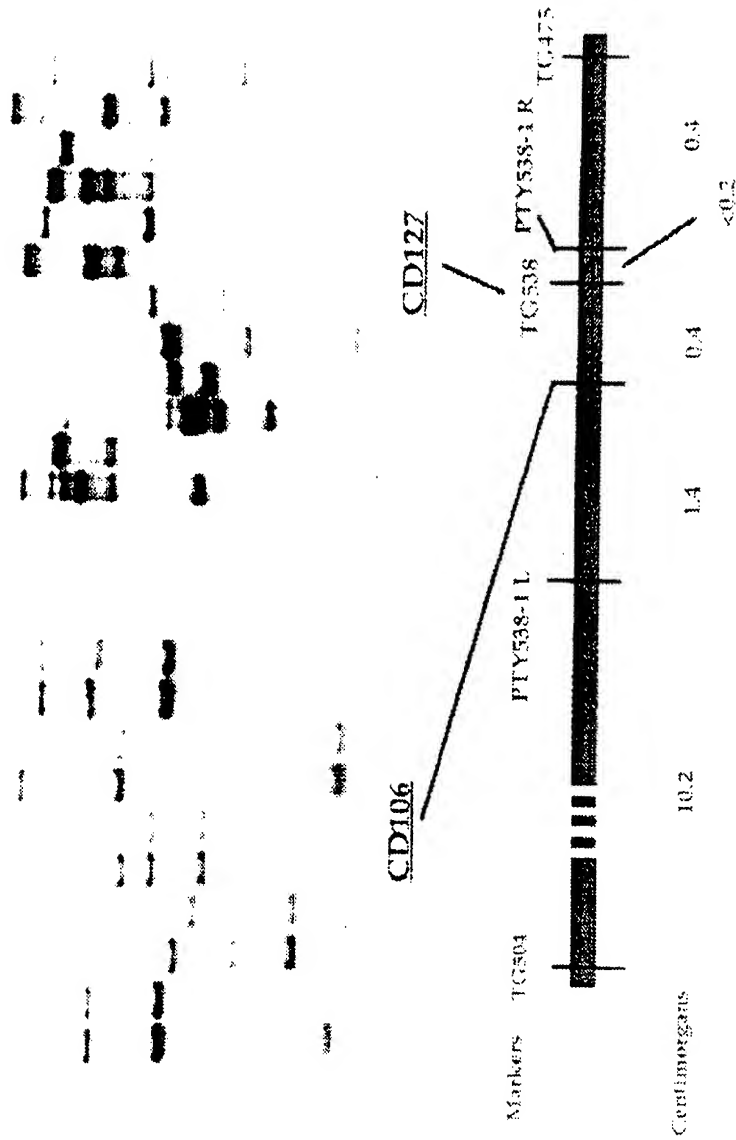
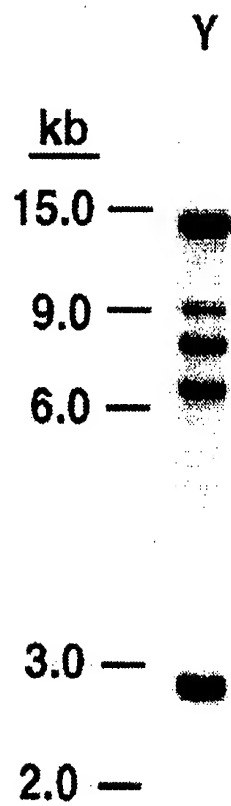
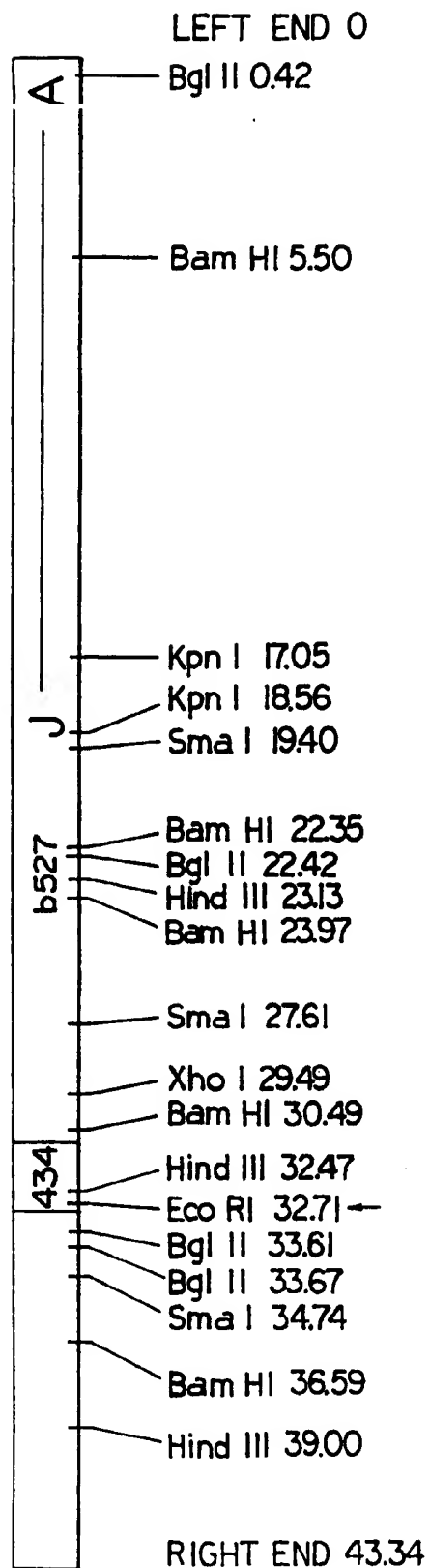


FIG.4

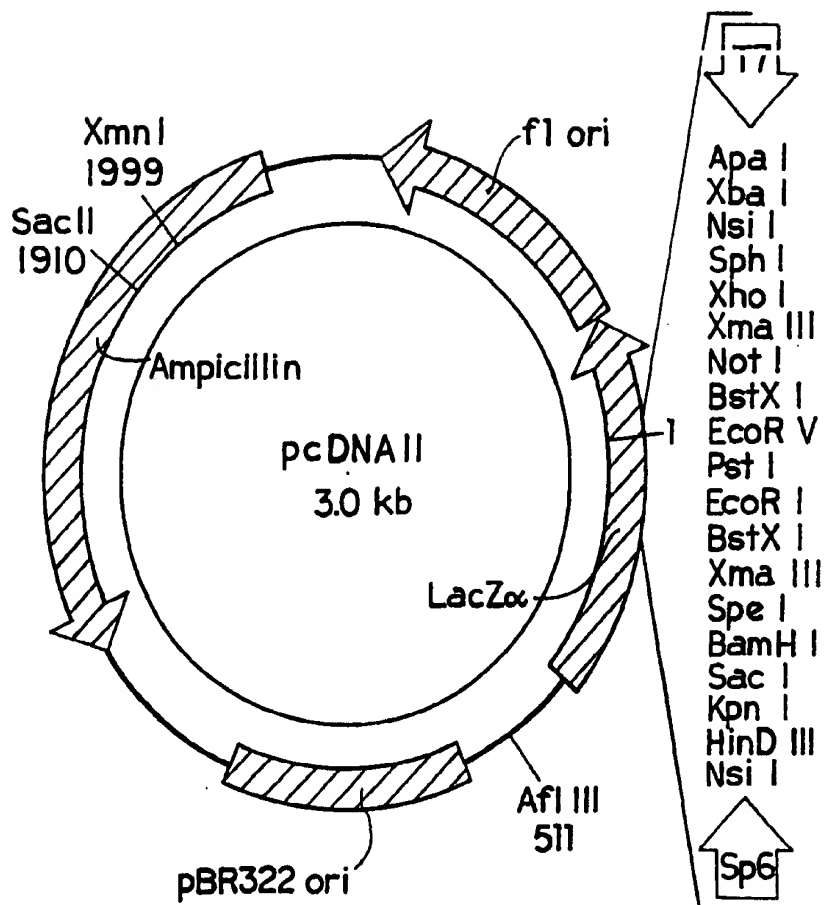
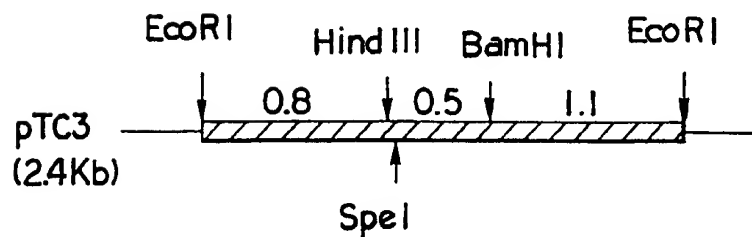
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FIG.5

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MAP OF LAMBDA
GT10 VECTORFIG.6

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FIG.7FIG.8

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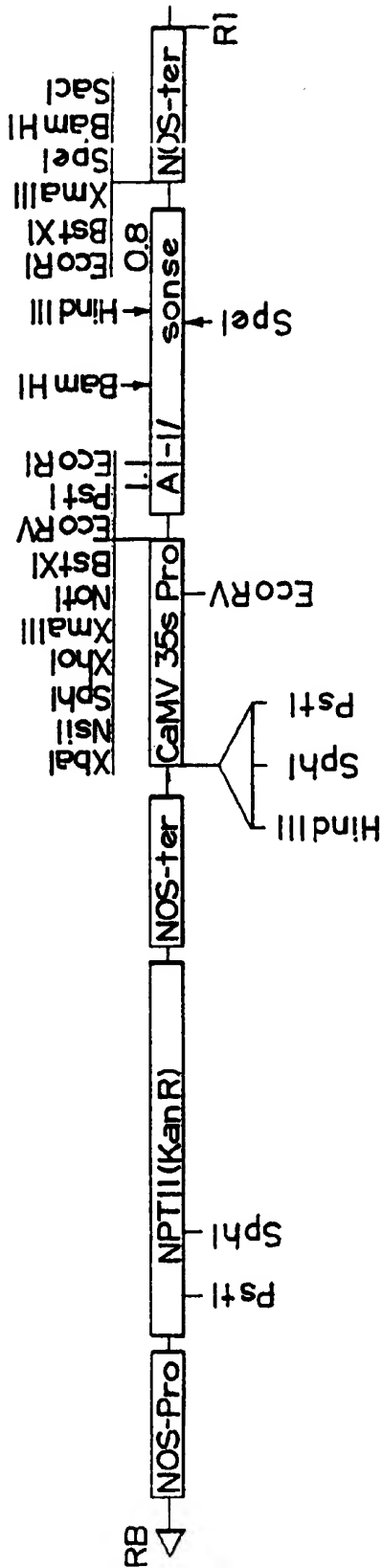


FIG. 9

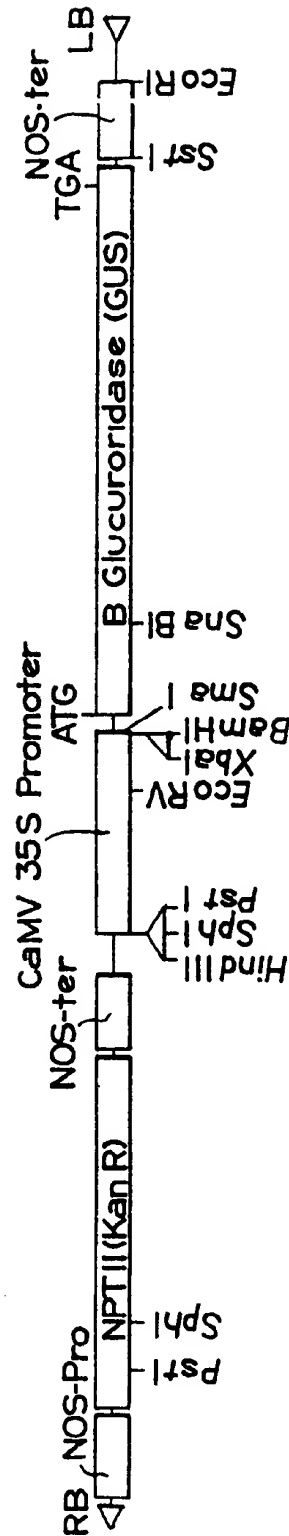
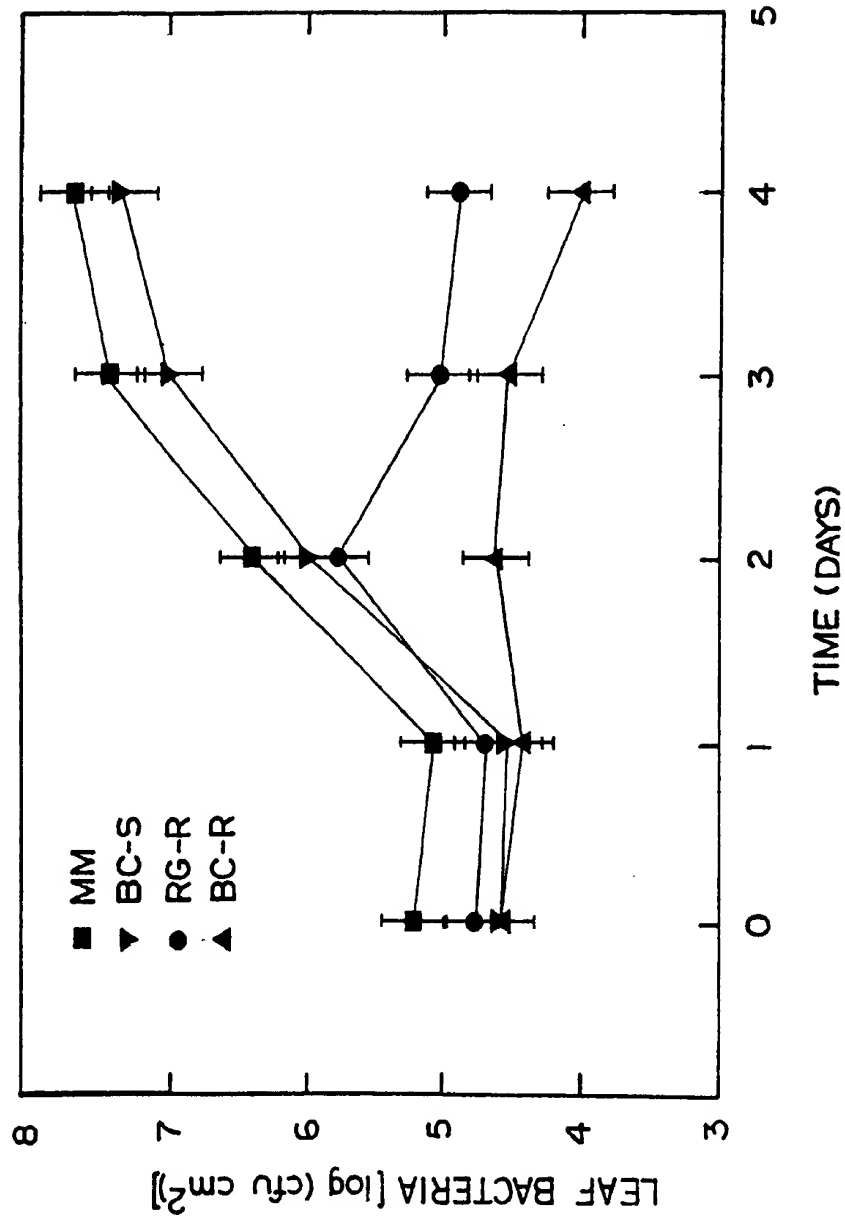
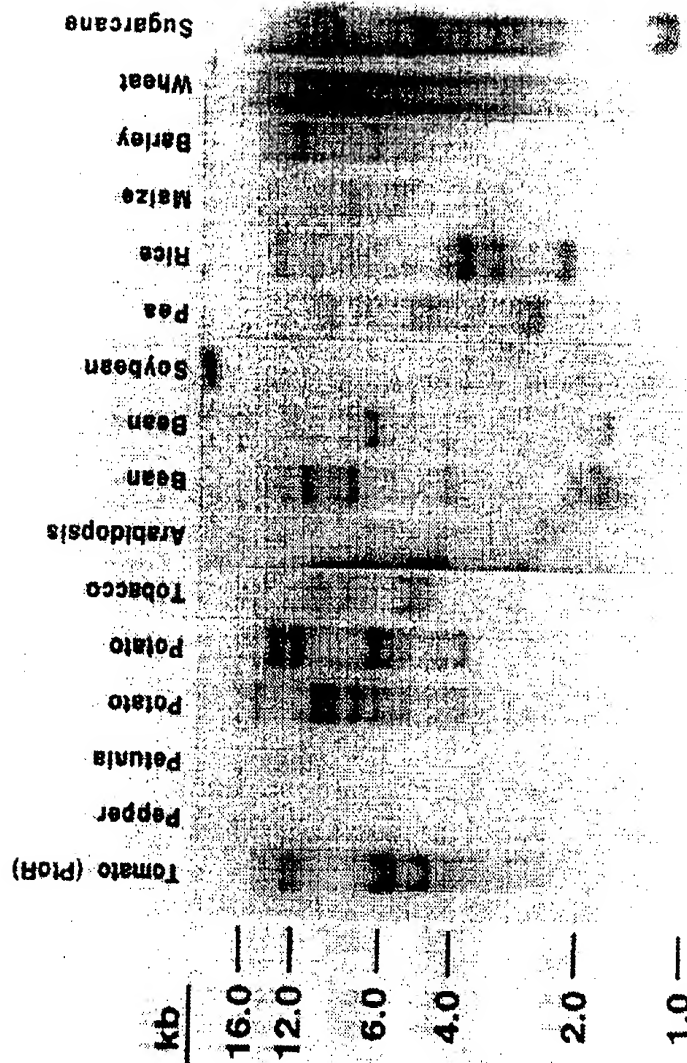


FIG. 10

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FIG. 11

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FIG. 12

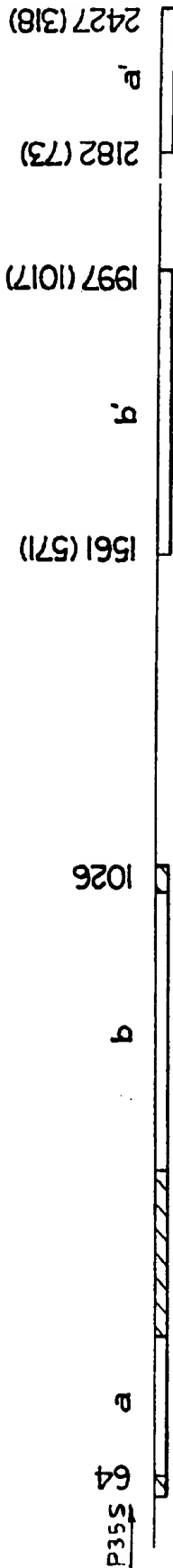


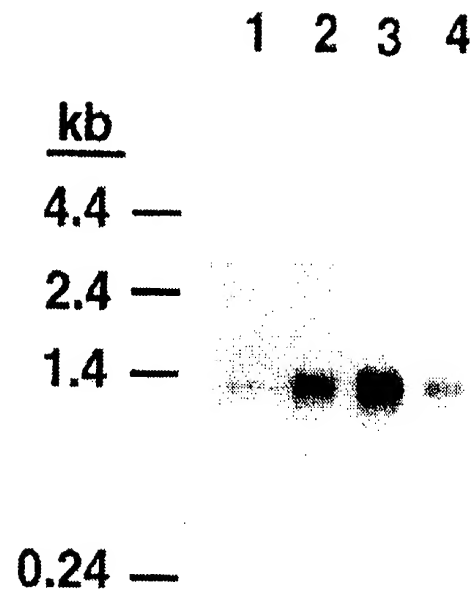
FIG.13a

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MGSKYSKATN	SINDALSSSY	LVPFESYRVP	IVDLEEATNN	FDHKFLIGHG	VFGKVYKGV	60
RDGAKVALKR	RTPESSQGI	EFETEITLS	FCRHPHLVSL	IGFCDERNEM	ILYKYMENG	120
NLKRHLYGSD	LPTMSMSWEQ	RLEICIGAAR	GLHTLHTRAI	IHRDVKSINI	LLDENFVPKI	180
TDFGISKKGT	ELDQTHLSTV	VKGTGLYIDP	EYFIKGRLTE	KSDVYSFGVV	LFEVLCARSA	240
IVQSLPREMV	NLAEWAVESH	NNGQLEQIVD	PNLADKIRPE	SLRKFGDTAV	KCLALSSEDR	300
PSMGDVLWKL	EYALRLQESV	I				321

FIG.13b

12/12

FIG. 14

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/09436

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/60.1, 70.1, 172.2, 124, 240.4, 252.2, 252.3, 320.1, 550/330, 550/23.2, 23.6; 800/205

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
NONEElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
NONE**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES USA, Volume 88, issued October 1991, J. C. Stein et al, "Molecular Cloning of a Putative Receptor Protein Kinase Gene Encoded at the Self-Incompatibility Locus of <i>Brassica oleracea</i> ", pages 8816-8820, see pages 8816-8817.	1-2, 7-8, 10-13, 15, 18, 19, 21-27, 29-30
Y	MOLECULAR AND GENERAL GENETICS, Volume 233, No. 1, issued 1992, G. B. Martin et al, "Construction of a Yeast Artificial Chromosome Library of Tomato and Identification of Cloned Segments Linked to Two Disease Resistance Loci", pages 25-32, see pages 30-31.	3-12, 14, 16-17, 20

☒ Further documents are listed in the continuation of Box C.
 ☐ See patent family annex.

Special categories of cited documents:		T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A	document defining the general state of the art which is not considered to be of particular relevance		
E	earlier document published on or after the international filing date	X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
L	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
O	document referring to an oral disclosure, use, exhibition or other means		
P	document published prior to the international filing date but later than the priority date claimed	A	document member of the same patent family

Date of the actual completion of the international search

13 NOVEMBER 1994

Date of mailing of the international search report

12 DEC 1994

 Name and mailing address of the ISA/US
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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/09436

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES USA, Volume 88, issued March 1991, G. B. Martin et al, "Rapid Identification of Markers Linked to a <i>Pseudomonas</i> Resistance Gene in Tomato by Using Random Primers and Near-Isogenic Lines", pages 2336-2340, see pages 2336 and 2338.	3-12, 14, 16-17, 20
Y	SCIENCE, Volume 245, issued 08 September 1989, J. M. Rommens et al, "Identification of the Cystic Fibrosis Gene: Chromosome Walking and Jumping", pages 1059-1065, see page 1063.	3-5, 7-12, 14, 16-17, 20
Y	BIO/TECHNOLOGY, Volume 6, issued April 1988, R. S. Nelson et al, "Virus Tolerance, Plant Growth, and Field Performance of Transgenic Tomato Plants Expressing Coat Protein from Tobacco Mosaic Virus," pages 403-409, see page 403.	28, 33
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES USA, Volume 84, issued October 1987, A. P. Bidwai et al, "Bacterial Phytotoxin, Syringomycin, Induces a Protein Kinase-Mediated Phosphorylation of Red Beet Plasma Membrane Polypeptides", pages 6755-6759, see pages 6755 and 6757.	10, 31-32, 34-40
Y	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES USA, Volume 88, issued October 1991, G. Felix et al, "Rapid Changes of Protein Phosphorylation are Involved in Transduction of the Elicitor Signal in Plant Cells", pages 8831-8834, see page 8831.	10, 31-32, 34-40

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/09436

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

A01H 1/04; C07H 21/04; C07K 14/00; C12N 5/00, 15/00; C12P 21/00, 21/04; C12R 1:41

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

435/69.1, 70.1, 172.3, 194, 240.4, 252.2, 252.3, 320.1; 530/350; 536/23.2, 23.6; 800/205